

# *ZytoFast* human Ig-kappa/Ig-lambda CISH Kit

**REF** T-1005-40  $\nabla_{\Sigma}$  40

**REF** T-1005-10  $\nabla_{\Sigma}$  10

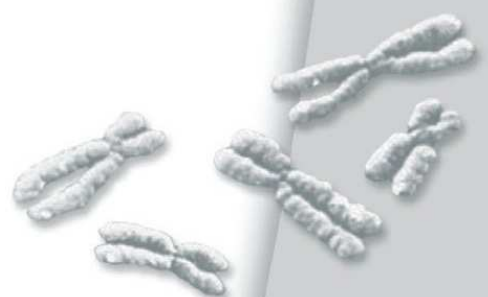
For the simultaneous detection of  
Ig-kappa ( $\kappa$ ) and Ig-lambda ( $\lambda$ ) mRNA  
by chromogenic *in situ* hybridization (CISH)



**IVD**

In vitro diagnostic medical device

according to EU directive 98/79/EC



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## 1. Scope of Application

The ZytoFast human Ig-kappa/Ig-lambda CISH Kit is designed to be used for the simultaneous detection of Ig-kappa ( $\kappa$ ) and Ig-lambda ( $\lambda$ ) mRNA in either formalin-fixed, paraffin-embedded tissue or cell samples by chromogenic *in situ* hybridization (CISH).

Interpretation of results must be made within the context of the patient's clinical history with respect to further clinical and pathologic data of patient by a qualified pathologist.

## 2. Basic Principles

The presence of certain nucleic acid sequences in cells or tissue can be detected with *in situ* hybridization using labeled DNA probes. The hybridization results in duplex formation of sequences present in the test object and the specific probe.

The ZytoFast human Ig-kappa/Ig-lambda CISH Kit uses the ZytoFast human Ig-kappa/Ig-lambda Probe (PF22). The probe contains digoxigenin-labeled oligonucleotides which target Ig-kappa ( $\kappa$ ) mRNA and biotin-labeled oligonucleotides which target Ig-lambda ( $\lambda$ ) mRNA.

Duplex formation of the digoxigenin-labeled probe can be visualized indirectly using a HRP-conjugated anti-digoxigenin antibody. The enzymatic reaction of AEC (3-amino-9-ethylcarbazole) leads to the formation of strong red signals. Duplex formation of the biotin-labeled probe can be visualized using AP-conjugated streptavidin. The enzymatic reaction of NBT/BCIP (Nitro blue tetrazolium chloride/5-Bromo-4-chloro-3-indolyl phosphate) leads to the formation of strong blue-violet signals that can be visualized by light microscopy at a 10-20x dry lens.

### 3. Safety Precautions and Disposal

- ✓ Read the operating instructions prior to use!
- ✓ Do not use the reagents after the expiry date has been reached!
- ✓ Avoid any cross-contamination and micro-bacterial contamination of the reagents!
- ✓ Some of the system components contain substances (in low concentrations and volumes) that are harmful to health. Avoid any direct contact with the reagents. Take appropriate protective measures (use disposable gloves, protective glasses, and lab garments)!
- ✓ If reagents come into contact with skin, rinse skin immediately with copious quantities of water!
- ✓ Never pipet solutions with your mouth!
- ✓ The disposal of reagents must be carried out in accordance with local regulations!
- ✓ A material safety data sheet is available on request for the professional user!

## 4. The ZytoFast human Ig-kappa/Ig-lambda CISH Kit

### 4.1 Components

The kit is made up of the following components:

Code	Component	Quantity		Container
		40 $\nabla \Sigma$	10	
ES1	<u>Pepsin Solution</u>	4 ml	1 ml	Dropper bottle, white cap
PF22	<u>ZytoFast</u> human Ig-kappa/Ig-lambda Probe	0.4 ml	0.1 ml	Reaction vessel, red lid
PF6	<u>ZytoFast</u> RNA (+) Control Probe	0.1 ml	0.1 ml	Reaction vessel, blue lid
PF7	<u>ZytoFast</u> RNA (-) Control Probe	0.1 ml	0.1 ml	Reaction vessel, white lid
AB8	<u>HRP-anti-Digoxigenin</u>	4 ml	1 ml	Dropper bottle, green cap
AB9	<u>AP-Streptavidin</u>	4 ml	1 ml	Dropper bottle, yellow cap
SB4	<u>NBT/BCIP</u>	4 ml	1 ml	Dropper bottle, blue cap
SB5	<u>AEC Solution</u>	4 ml	1 ml	Dropper bottle, red cap
WB5	<u>20x Wash Buffer TBS</u>	2x 50 ml	50 ml	Screw-cap bottle
	Instruction manual	1	1	

**T-1005-40 (40 tests):** Components **(ES1)**, **(PF22)**, **(AB8)**, **(AB9)**, **(SB4)**, and **(SB5)** are sufficient for 40 reactions. Components **(PF6)** and **(PF7)** are sufficient for 10 reactions. Component **(WB5)** is sufficient for 28 staining jars of 70 ml each.

**T-1005-10 (10 tests):** Components **(ES1)**, **(PF22)**, **(AB8)**, **(AB9)**, **(SB4)**, and **(SB5)** are sufficient for 10 reactions. Components **(PF6)** and **(PF7)** are sufficient for 10 reactions. Component **(WB5)** is sufficient for 14 staining jars of 70 ml each.

### 4.2 Storage and Shelf Life

The components of the kit must be stored at 2...8°C.

If these storage conditions are followed, the kit will function, without loss of performance, at least until the expiry date printed on the label.

## 4.3 Test Material

The ZytoFast human Ig-kappa/Ig-lambda CISH Kit has been optimized for the use with formalin-fixed, paraffin-embedded tissue and cell samples. When test material is used that has been fixed or embedded in a different manner (e.g. methanol/glacial-acetic-acid-fixed cells or blood smears) the test protocol may need to be adapted by the user. Our specialists are available to support you whenever adjustments are necessary.

We recommend the following tissue preparation:

- ✓ Fixation in 10% neutrally buffered formalin for 24 h at RT  
*In order to achieve optimum and uniform fixation and paraffin embedding, the sample size should not exceed 0.5 cm<sup>3</sup>.*
- ✓ Standard processing and paraffin embedding  
*Use premium quality paraffin. Infiltration and embedding should be carried out at temperatures lower than 65°C.*
- ✓ Prepare 2-5 µm microtome sections  
*Draw up the sections onto silane-coated or adhesion slides (e.g. HistoBond®) and fix for 2-16 h at 50-60°C.*

## 4.4 Additional Materials

The following reagents, materials, and equipment are not included in the kit:

Reagents and materials

- Adhesive pistol, including hot adhesive, or rubber cement (Fixogum)
- Ethanol 100%, denatured
- Deionized or distilled water
- Xylene
- Counterstain
- Aqueous mounting medium
- Hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) 30%
- Methanol 100%

Equipment

- Water bath (55°C)
- Hot plate
- Hybridization oven (heating oven)
- Staining jars, 50-80 ml
- Humidity chamber
- Pipet (10 µl)

- Coverslips (22 mm x 22 mm, 24 mm x 32 mm)
- Light microscope

## 4.5 Important Information

The following should be kept in mind:

- ✓ The tissue and cell sections must not be allowed to dry during the hybridization and washing steps!
- ✓ The temperature for denaturing and washing, described in the protocol, should be used as a guide. Dependent upon the age and the fixation step of the sample material, an increase or decrease in temperature of the denaturing or wash steps can lead to better hybridization results!
- ✓ Do not wash slides hybridized with ZytoFast RNA (+) Control Probe (PF6) at 55°C as this will reduce signal intensity!

## 5. The ZytoFast human Ig-kappa/Ig-lambda CISH Kit Protocol

### 5.1 Preparatory Steps

*Preparation of 3% H<sub>2</sub>O<sub>2</sub>:* Dilute 1 part of 30% H<sub>2</sub>O<sub>2</sub> with 9 parts of 100% methanol.

*Preparation of 1x Wash Buffer TBS:* Dilute 1 part of 20x Wash Buffer TBS (WB5) in 19 parts deionized or distilled water. Diluted 1x Wash Buffer TBS lasts for one week when stored at 2-8°C.

*1x Wash Buffer TBS:* Prepare one staining jar with 1x Wash Buffer TBS (prepared using **WB5**) and heat in a water bath to 55°C.

ZytoFast human Ig-kappa/Ig-lambda Probe (PF22), ZytoFast RNA (+) Control Probe (PF6), ZytoFast RNA (-) Control Probe (PF7): Bring to hybridization temperature before use.

Pepsin Solution (ES1), HRP-anti-Digoxigenin (AB8), AP-Streptavidin (AB9), NBT/BCIP (SB4), AEC Solution (SB5): Bring to room temperature before use.



## 5.2 Pretreatment (Dewax/Proteolysis)

1. Incubate slides for 10 min at 70°C (e.g. on a hot plate)
2. Incubate slides for 2x 5 min in xylene
3. Incubate 3x 3 min in 100% ethanol

*Alternatively, dewaxing protocols routinely used in immunohistochemistry procedures, e.g. 2x 15 min xylene, 2x 5 min 100% ethanol, 2x 5 min 96% ethanol, 1x 5 min 70% ethanol, can be used.*

4. Incubate slides for 10 min in 3% H<sub>2</sub>O<sub>2</sub>
5. Wash 3x 1 min in deionized or distilled water and drain off or blot off the water
6. Apply (dropwise) Pepsin Solution (ES1) to the tissue/cell section and incubate for 20-30 min at 37°C in a humidity chamber

*Depending on multiple factors, e.g. nature and duration of fixing, thickness of sections, and nature of tissue/cells, different incubation times may be required. As an incubation guideline, we recommend an incubation time of 10-30 min for tissue samples and 5-10 min for cell samples. As a general rule, we recommend to ascertain the optimum time for proteolysis in pre-tests.*

7. Immerse slides in deionized or distilled water and drain off or blot off the water

Air dry sections

## 5.3 Denaturation and Hybridization

1. Vortex the ZytoFast human Ig-kappa/Ig-lambda Probe (PF22) and pipette 10 µl each onto individual samples

*Positive control: Pipet 10 µl ZytoFast RNA (+) Control Probe (PF6) onto an additional slide with the test material.*

*Negative control: Pipet 10 µl ZytoFast RNA (-) Control Probe (PF7) onto an additional slide with the test material.*

*Distribute dropwise on the whole target area to avoid local concentration of probe. Alternatively, add probe to the center of a coverslip and place coverslip upside down on target area.*

2. Avoiding trapped bubbles, cover the samples with a coverslip (22 mm x 22 mm). Seal the coverslip, e.g. with a layer of hot glue from an adhesive pistol or with rubber cement
3. Denature the slides at 75°C for 5 min, e.g. on a hot plate
4. Transfer the slides to a humidity chamber and hybridize for 2 h at 55°C (e.g. in a hybridization oven)

*It is essential that the tissue/cell samples do not dry out during the hybridization step.*

## 5.4 Post-Hybridization and Detection

1. Carefully remove the rubber cement or glue
2. Remove the coverslip by submerging in 1x Wash Buffer TBS (prepared using **WB5**) at RT for 5 min

*If only the sealant can be removed and the coverslip remains on the slide, the coverslip can be rinsed off with the wash buffer during this washing step. In such cases, this wash step should be extended to 10 min.*

3. Wash 5 min in 1x Wash Buffer TBS (prepared using **WB5**) at 55°C

*Do **NOT** perform this step on slides hybridized with ZytoFast RNA (+) Control Probe (PF6) as this will reduce signal intensity!*

*1x Wash Buffer TBS must be sufficiently prewarmed. Check with a thermometer if necessary.*

4. Wash 5 min in 1x Wash Buffer TBS (prepared using **WB5**) at RT
5. Apply HRP-anti-Digoxigenin (AB8) dropwise (3-4 drops per slide) to the slides and incubate for 30 min at 37°C in a humidity chamber

**6.** Wash 2x 2 min in 1x Wash Buffer TBS (prepared using **WB5**) and 1x 2 min in deionized or distilled water

**7.** Apply AEC Solution (**SB5**) dropwise (3-4 drops per slide) to the slides and incubate for 20-40 min at 37°C in a humidity chamber

*It is recommended that you check the color development in intervals of approx. 5-10 min using a microscope.*

**8.** Wash 3x 2 min in deionized or distilled water

**9.** Apply AP-Streptavidin (**AB9**) dropwise (3-4 drops per slide) to the slides and incubate for 30 min at 37°C in a humidity chamber

**10.** Wash 2x 2 min in 1x Wash Buffer TBS (prepared using **WB5**) and 1x 2 min in deionized or distilled water

**11.** Apply NBT/BCIP (**SB4**) dropwise (3-4 drops per slide) to the slides and incubate for 20-40 min at 37°C in a humidity chamber

*It is recommended that you check the color development in intervals of approx. 5-10 min using a microscope.*

**12.** Wash 3x 2 min in deionized or distilled water

**13.** Cover the sections

*We recommend that the sections should be embedded in an aqueous embedding medium.*

**14.** Evaluation of the sample material is carried out by light microscopy

## 6. Interpretation of Results

The ZytoFast human Ig-kappa/Ig-lambda CISH Kit procedure causes red and/or blue-violet colored precipitates within the cells targeted by the ZytoFast human Ig-kappa/Ig-lambda Probe (PF22), which can be clearly distinguished from the background.

A positive reactivity for Ig-kappa ( $\kappa$ ) mRNA in the target cells is indicated by red cytoplasmic staining.

A positive reactivity for Ig-lambda ( $\lambda$ ) mRNA in the target cells is indicated by blue-violet cytoplasmic staining.

Visualization of signals should be performed by light microscopy using a 10x or 20x objective. For signal evaluation, necrotic, degenerated or over-digested cells should be avoided as these cells often stain nonspecifically.

In order to judge the specificity of the hybridization signals and to confirm the correct performance of the method, any hybridization should be accompanied by controls. We recommend using at least one control sample containing both true positive and negative staining cells.

A negative or unspecific result can be caused by multiple factors. For troubleshooting, please refer to chapter 8.

The ZytoFast RNA (+) Control Probe (PF6) consists of poly-dT oligonucleotides targeting the poly(A) tails of mRNAs. Strong blue-violet hybridization signals within the cytoplasm of cells verify the integrity of cellular mRNA in specimens.

The ZytoFast RNA (-) Control Probe (PF7) consists of a set of random sequence oligonucleotides with GC contents of 40-70% without known consensus to any naturally occurring sequences. This probe should not result in positive staining signals and is to be used to assess the unspecific background staining within specimens.

## 7. Literature

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## 8. Problems and Possible Causes

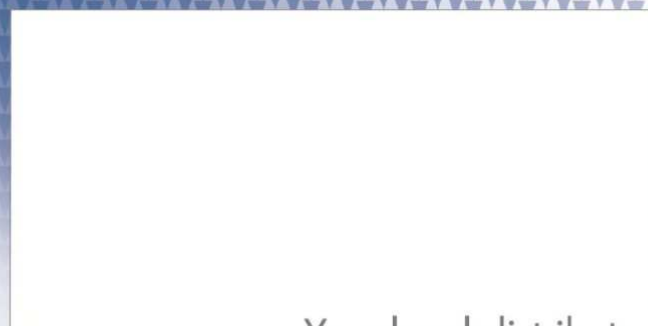
Any deviation from the operating instructions can lead to inferior staining results or to no staining at all.

Problem	Possible cause	Action
Streaks on the slide after stopping the pepsin treatment	Precipitation	Wash section immediately in deionized or distilled water
Weak signal or no signal at all	No target sequences present or target sequences below detection limit	Use controls
	Low amount of target sequences	Extension of hybridization and antibody incubation times
	Cell or tissue sample has not been properly fixed	Optimization of fixing time
	Proteolytic pretreatment not carried out properly	Optimization of incubation time
	Denaturation temperature not correct	Check temperature; increase or decrease if necessary
	Hybridization temperature not correct	Check temperature
	Hybridization time too short	Extension of hybridization time
	Incubation with chromogenic substrate too short	Extension of incubation time
Uneven and in some parts only very light staining	Incomplete dewaxing	Use fresh solutions; check length of dewaxing times
Cross hybridization signals; strong background staining	Incomplete dewaxing	Use fresh solutions; check length of dewaxing times
	Probe volume per area too high	Reduce probe volume per section/area, distribute probe dropwise to avoid local concentration
	Proteolytic pretreatment too strong	Optimization of incubation time
	Dehydration of sections between the individual incubation steps	Prevent dehydration
	Washing temperature following hybridization too low	Check temperature
Section floats off the slide	Proteolytic pretreatment too strong	Shortening of incubation time
	Unsuitable slide coating	Use appropriate slides





ZytoVision GmbH · Fischkai 1  
D - 27572 Bremerhaven · Germany  
Phone: +49 (0)471/4832 - 300  
Fax: +49 (0)471/4832 - 509  
[www.zytovision.com](http://www.zytovision.com)  
[info@zytovision.com](mailto:info@zytovision.com)



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