

## Instructions For Use UAP-IFU

Rev. Date: 10/29/03

Revision: 2

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 P.O. Box 3286 - Logan, Utah 84323, U.S.A. - Tel. (800) 729-8350 - Fax (435) 755-0015 - [www.scytek.com](http://www.scytek.com)

# UltraTek Alk-Phos Anti-Polyvalent Lab Pack

**Species of Origin:** Goat  
**Antigen Specificity:** Anti-Polyvalent (Mouse, Rat, Rabbit and Guinea Pig.  
**Preadsorbed Against:** Human  
**Enzyme Conjugate:** Alkaline Phosphatase  
**Chromogen Substrate:** None Provided

**Uses/Limitations:** For Research Use Only.  
 Do not use past expiration date.  
 For immunohistochemical studies.

### Availability:

Item #	Volume
UAP125	125ml Super Block, 125ml UltraTek Anti-Polyvalent, 125ml UltraTek Alk-Phos.
UAP500	500ml Super Block, 500ml UltraTek Anti-Polyvalent, 500ml UltraTek Alk-Phos.
UAP999	1000ml Super Block, 1000ml UltraTek Anti-Polyvalent, 1000ml UltraTek Alk-Phos.

**Storage:** 2-8° Centigrade.

### Procedure:

1. Deparaffinize and rehydrate tissue section.
2. Wash 2 times in Tris Buffered Saline + Tween 20 (20X) pH 7.4 (catalog # TBT500).
3. If required, incubate tissue in digestive enzyme (catalog # PSS060 or TSS155) or Citrate Plus (catalog # CPL500).
4. Wash 3 times in Tris Buffered Saline + Tween 20 (20X) pH 7.4 (catalog # TBT500).
5. Apply Super Block and incubate for 5 minutes at room temperature to block nonspecific background staining. Note: Do not exceed 10 minutes or there may be a reduction in desired stain.
6. Wash 1 time in Tris Buffered Saline + Tween 20 (20X) pH 7.4 (catalog # TBT500).
7. Apply primary antibody and incubate according to manufacturer's protocol.
8. Wash 3 times in Tris Buffered Saline + Tween 20 (20X) pH 7.4 (catalog # TBT500).
9. Apply UltraTek Anti-Polyvalent (yellow solution), and incubate for 10 minutes at room temperature.
10. Wash 3 times in Tris Buffered Saline + Tween 20 (20X) pH 7.4 (catalog # TBT500).
11. Apply UltraTek Alk-Phos (red solution), and incubate for 10 minutes at room temperature.

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12. Wash 3 times in Tris Buffered Saline + Tween 20 (20X) pH 7.4 (catalog # TBT500).
13. Apply chromogen intended for use with Alkaline Phosphatase and incubate as desired.
14. For optimal results counterstain using Hematoxylin for Automation (catalog # HAQ500).
15. Coverslip using Aqueous mounting media (catalog # AMT030 or PMT030).

### Troubleshooting Guide

#### **Overstaining:**

1. Concentration of the primary antibody was too high or the incubation time was too long.
2. Temperature during incubation was too high.
3. Incubation time with link antibody or streptavidin/enzyme label was too long.

#### **Nonspecific Background Staining:**

1. Rinsing between steps was inadequate.
2. Tissue was allowed to dry with reagents on.
3. Folds in tissue trapped reagents.
4. Tissue contains endogenous alkaline phosphatase.
5. Tissue contains endogenous biotin.
6. Antigen migrated in tissue.
7. Excessive tissue adhesive on slides.
8. Inadequate blocking with protein block.

#### **Weak Staining:**

1. Primary antibody concentration was too low or incubation time was too short.
2. Reagents are past their expiration date.
3. Inadequate removal of wash water between steps, resulting in dilution of reagents.
4. Counterstain or mounting media were incompatible and dissolved the chromogen reaction product.
5. Room temperature was excessively cool.
6. The primary antibody does not recognize an antigen that survives fixation and embedding in high enough amounts.
7. Excessive incubation with protein block (Super Block).

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### No Staining:

1. Steps were inadvertently left out.
2. There is no antigen in the tissue.
3. The primary antibody is not of mouse, rat, rabbit or guinea pig origin.
4. Chromogenic substrate has been replaced with another that is not intended for use with alkaline phosphatase.

**Precautions:** Handle with care and dispose of according to all regulations.