

# FITkit®

## **FITkit® Hev b 1 CAT K3-350-020**

Instructions for use of FITkit® Hev b 1 in  
quantitative determination of Hev b 1 in  
natural rubber latex products

Icosagen is an innovative biotech company providing technologies, products and services in the field of molecular and cellular biology, biochemistry and immunoanalysis.

Proprietary technologies are used combined with extensive process know-how and partnership with academic and industrial groups.



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## 1. Introduction

Products containing natural rubber latex (NRL) from the rubber tree *Hevea brasiliensis* are widely used due to the economical price and advantageous processing properties of natural rubber, although adverse reactions against a number of allergenic proteins contained in the NRL are well known and documented. The NRL-containing products used by, e.g., healthcare personnel, such as surgical gloves and various other devices (like catheters, tubes, masks, etc.) contribute to the major portion of these adverse reactions. In healthcare, the NRL-based medical devices exhibit a potential danger not only to the personnel but also to the patients undergoing an examination or a surgery. Additionally, even the general population comes into daily contact with diverse NRL-containing products, such as household gloves, condoms and balloons, manufactured by the dipping procedure, and also with tubes, tires, erasers and the like.

Currently, latex allergy is recognized as a serious world-wide health problem: up to 15 % of health care workers and approximately 1 % of the entire population are allergic to NRL. The clinical manifestations of latex allergy range from mild contact urticaria to fatal anaphylaxis and the seriousness of the condition is accentuated by the fact that the first sign of sensitization can manifest as a life-threatening reaction.

Latex allergens are proteins or polypeptides eluting from the manufactured products upon contact with skin, mucous membranes or other tissues. According to the current allergen nomenclature system maintained by the International Union of Immunological Societies (IUIS) under the WHO, thirteen latex allergens, which have been characterized at the primary structure level and are contained in the official allergen list, are named as Hev b 1, Hev b 2, Hev b 3, Hev b 4, Hev b 5, Hev b 6 (6.01, 6.02, 6.03), Hev b 7, Hev b 8, Hev b 9, Hev b 10, Hev b 11, Hev b 12 and Hev b 13. Not all previously listed allergens are able to come through the production process. At present, four of these allergens (Hev b 1, Hev b 3, Hev b 5 and Hev b 6.02) have been mostly detected in manufactured latex products. According to the current literature, the sum quantity of these four allergens is highly dominant in determining the allergenic potential of NRL products.

FITkit® Hev b 1 test is the first commercial quantitative test to measure Hev b 1 immunologically in NRL products. By the use of specific monoclonal antibodies, sensitivity and specificity stay high even in the presence of other proteins or chemical substances derived from the manufacturing process of NRL products.

FITkit® Hev 1 test is produced in accordance with ASTM International standard D7427-08 ([www.astm.org/Standards/D7427.htm](http://www.astm.org/Standards/D7427.htm))

## 2. Principle of method

FITkit® Hev b 1 test is based on the immunoenzymetric assay technique. Microtiter wells are coated with Hev b 1-specific monoclonal capture antibody that during the first incubation binds Hev b 1 molecules from the sample extract. After incubation, unbound material is removed by washing the wells. During the second incubation, horseradish peroxidase (HRP) labelled Hev b 1-specific monoclonal detection antibody binds to a different epitope of Hev b 1 molecules. As such, the two antibodies are a matched pair that forms a "sandwich" around the target Hev b 1 allergen. After washing, HRP substrate is added and the intensity of the colour produced is directly proportional to the Hev b 1 concentration of the sample.

## 3. Contents of kit

The kit contains the reagents listed below, sufficient for 96 wells.

### 3.1 FITkit® Hev b 1 Microwell Plate

Cat F3-301-001

96 wells coated with mouse monoclonal Hev b 1 antibody, packed in a aluminium vacuum bag. The microwell plate is ready for use.

### 3.2 FITkit® Hev b 1 Assay Buffer, 15 mL

Cat F3-301-041

The Assay Buffer contains phosphate, sodium chloride, EDTA, bovine plasma albumin (BPLA), detergent and preservative Proclin 300®. Ready for use. Coloured red.

### 3.3 FITkit® Hev b 1 Calibrators

Cat F3-301-031...036

Each vial contains 0.5 mL Hev b 1 calibrator in a stabilized buffer. The calibration is based on the analysis of Hev b 1 in reversed phase chromatography and N-terminal sequencing. The calibrator values are 0, 10, 50, 200, 500 and 1000 µg/L. Ready for use.

### 3.4 FITkit® Hev b 1 Control

Cat F3-301-081

The control is made from field latex in a stabilized solution containing BPLA, detergents and preservatives. Reconstitute the lyophilized control with 500 µL of distilled water.



### 3.5 FITkit® Hev b 1 Enzyme Conjugate, 15 mL

Cat F3-301-016

Mouse monoclonal Hev b 1 antibody conjugated to horse radish peroxidase (HRP) in a buffered solution containing stabilizers, BPLA, detergent and preservative Proclin 300®. Ready for use.

### 3.6 FITkit® PBS Wash Concentrate, 50 mL

Cat F3-300-042

Before use dilute to 500 mL (1:10) with distilled water. Occasionally crystals may be present at +2...+8 °C, but they dissolve upon dilution and at room temperature.

### 3.7 FITkit® HRP Substrate Solution, 15 mL

Cat F3-300-043

ABTS (2,2'-azino-di[3-ethyl-benzthiazoline-6-sulphonate]) Peroxidase Substrate. Ready for use. Colourless to light green.

### 3.8 FITkit® Stopping Solution, 15 mL

Cat F3-300-044

1 % Sodium dodecyl sulphate (SDS). SDS precipitates at low temperatures, but redissolves upon warming to room temperature. Ready for use.

## 4. Storage conditions

The kit should be stored at +2...+8 °C.

The unopened kit is stable until the expiry date, printed on the kit label. The expiry date of each unopened component is printed on the label of the individual component.

Once opened, the microwell plate and liquid components are stable for eight weeks at +2...+8 °C.

After reconstitution, the control should be used during the same working day.

After dilution of the PBS Wash Concentrate, the washing solution is stable for eight weeks at room temperature (RT).

## 5. Materials and equipment required but not supplied

- Pipette with disposable plastic tips: 25 µL (for calibrators and samples), 500 µL (for the reconstitution of the control).
- Multichannel pipette with disposable plastic tips: 100 µL (assay buffer, enzyme conjugate, substrate, stopping solution).
- PBS (phosphate buffered saline) for sample preparation.
- Lid or sealing tape for microwell plate.
- Reagent troughs.
- Microwell plate shaker.
- Aspiration device or microwell washer.
- Photometer (microwell plate or strip reader), 414 nm or 405 nm.

## 6. Precautions and notes

- Wear only synthetic disposable gloves during the assay. Do not use any gloves or other materials containing natural rubber latex.
- Always use the polypropylene low protein binding vessels and tubes in every step. Do not use glass tubes because of the possible loss of proteins by adsorption to the inner surface of the tube walls.
- Protect the microwell plate from draught, strong light or direct sunlight during the test procedure.
- Each microwell plate should include a standard curve on the same plate and at the same time as control and samples.
- Careful aspiration of the washing solution is essential for good assay precision. It is recommended that the washing procedure mode is checked to get the best precision.
- Timing of the incubation steps is important to the performance of the assay. Pipetting of calibrators, control and samples should be done without interruption. Pipetting of the calibrators and samples should not exceed 10 minutes to avoid assay drift.
- Adding of substrate starts a kinetic reaction that is terminated by dispensing the stopping solution. Keep the incubation times for each well the same by adding the reagents at timed intervals.
- Absorbance values are stable for 60 minutes if protected from light.
- Microwell plate readers measure absorbance vertically. Do not touch the bottoms of the wells.
- A wavelength of 405 nm can be used if 414 nm is not available. Absorbances are slightly lower at 405 nm than at 414 nm.

## 7. Preparation of samples

### 7.1 A. For solid-state material:

Weigh the entire natural rubber product and record the total weight per product so that the allergen content can be later reported as micrograms of allergen per gram of product weight. Clearly non-NRL parts may be removed\*.

*\* If non-NRL parts were removed from the product, weigh, calculate and report only the analysed NRL part of the product. For example, if the product is a baby pacifier with a soft nipple and stiff plastic base, remove the plastic base, weigh, extract and analyse only pacifier nipple pieces without the plastic base. Report the result in µg/g with the additional claim that the result value is per partial product (pacifier nipple) not per entire product.*

**B.** For solutions- Proceed to Step 7.6.

**7.2** Cut the product into pieces using clean and completely dry scissors.

**7.3** Place the pieces of test specimen into an extraction vessel. Extract in PBS (phosphate buffered saline) using volume to weight ratio 5:1 (5mL PBS per 1g of NRL product). All product surfaces should be evenly exposed to PBS.

**7.4** Extract the specimen pieces at room temperature (21-23°C) for 2 hours on an end-over-end or an orbital shaker. The extraction vessel should be continuously rotated to ensure even exposure to the PBS. Avoid foaming.

**7.5** Remove the test specimen pieces from the extraction buffer.

**7.6** Centrifuge the supernatant 2000 g for 15 min at RT.

**7.7** Collect the supernatant liquid and assay immediately or store at +4°C up to 4 hours. If Hev b 1 determination is performed later, the samples should be kept frozen (-20°C). Avoid repeated freeze thaw cycles.

## 8. Procedure of test

### 8.1 Preparation of reagents and equipment

Allow all reagents to reach room temperature (21-23°C) before use. Reconstitute the control (Step p 8.3.2) and let it dissolve for 2 hours, mix gently (or vortex for 1-2 seconds).

Dilute the PBS Wash Concentrate to make washing solution (Step 8.3.3). Take the required number of microwell plate strips and place the remaining strips back into the aluminium vacuum bag. Close the bag tightly again.

### 8.2 Test procedure

**8.2.1** Dispense 100 µL of Assay Buffer into each well.

**8.2.2** Pipette 25 µL of calibrator, control and sample into appropriate wells in duplicate.

**8.2.3** Cover the microwell plate. Incubate the microwell plate for 60 minutes at room temperature on a microwell plate shaker (100-200 rpm).

**8.2.4** Aspirate and wash the wells 4 times with 300 µL of washing solution.

**8.2.5** Dispense 100 µL of Enzyme Conjugate into the wells in duplicate.

**8.2.6** Cover the microwell plate. Incubate the microwell plate for 30 minutes at room temperature on a microwell plate shaker (100-200 rpm).

**8.2.7** Aspirate and wash the wells 4 times with 300 µL of washing solution.

**8.2.8** Add 100 µL of HRP Substrate Solution at fixed time points into each well.

**8.2.9** Cover the microwell plate. Incubate the microwell plate for 15 minutes at room temperature on a microwell plate shaker (100-200 rpm).

**8.2.10** Stop the reaction by adding 100 µL of Stopping Solution into each well at the same fixed time points as in Step 8.2.8 so that exactly the same substrate reaction time is achieved. Shake the microwell plate for 1-2 minutes to mix the solutions.

**8.2.11** Measure the absorbance at 414 nm using a microwell plate or strip reader, preferably immediately but no more than 60 minutes after stopping the reaction. If the microwell plate is not read immediately, protect the microwell plate from light.

### 8.3 Summary - test procedure

#### Preparative steps

1.



Bring all reagents to room temperature.

2.



Reconstitute Control with 500  $\mu$ L of distilled water. Let it stand for 2 hours. Mark the wells to be used.

3.



Dilute 50 mL Wash Concentrate with 450 mL of distilled water.

#### Test procedure

4.



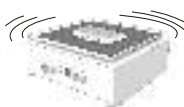
Dispense 100  $\mu$ L Assay Buffer

5.



Add 25  $\mu$ L calibrators, controls and product extracts into appropriate wells.

6.



Incubate for 60 min with shaking at RT.

7.



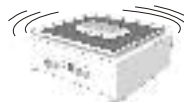
Wash 4 times.

8.



Dispense 100  $\mu$ L Enzyme Conjugate.

9.



Incubate for 30 min with shaking at RT.

10.



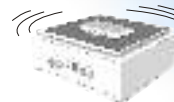
Wash 4 times.

11.



Dispense 100  $\mu$ L Substrate Solution.

12.



Incubate for 15 min with shaking at RT.

13.



Dispense 100  $\mu$ L Stopping Solution.

14.



Shake for 1-2 minutes and measure at 414 nm (or 405 nm). Fit the curve and read off results.

## 8.4 Calculation of results

Calculate the mean absorbance for each duplicate. Subtract blank values (Calibrator A) from the mean absorbances.

Plot the absorbances against the respective Hev b 1 concentrations on a log-log scale. Software that fits the standard curve can be used to calculate results of samples. Cubic spline fit type is recommended.

Read off the concentrations of the controls and samples (µg/L). Samples showing lower concentrations of Hev b 1 compared to Calibrator B (10 µg/L) are considered as undetectable. Samples showing higher concentrations of Hev b 1 compared to Calibrator F (1000 µg/L) must be diluted further to obtain result. If samples have been diluted, multiply the result with the dilution factor.

Report Hev b 1 protein content in microgram of Hev b 1 per gram (µg/g) of product. To convert Hev b 1 concentration from µg/L to µg/g, use following equation:

$$C1 (\mu\text{g/g}) = \frac{C2 (\mu\text{g/L}) * V1 (\text{L})}{W1 (\text{g})}$$

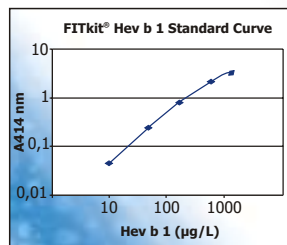
C1 - Hevb 1 content in (µg/g)

C2 - Hevb 1 concentration (µg/L)

V1 - Volume of PBS used for sample extraction (L)

W1 - Weight of the product (g) determined in Step 7.1.A

*Worksheet and standard curve of typical assay (Not to be used for calculation of actual test results)*



**FITkit® Hev b 1 test**

Wells	Identity	Conc. µg/L	A414 nm	A <sub>414 nm</sub> -blank	Conc. µg/L
A1-A2	Calibr. A	0	0,075		
B1-B2	Calibr. B	10	0,118	0,043	
C1-C2	Calibr. C	50	0,308	0,233	
D1-D2	Calibr. D	200	0,914	0,839	
E1-E2	Calibr. E	500	1,996	1,921	
F1-F2	Calibr. F	1000	2,993	2,918	
G1-G2	Sample 1	unknown	0,235	0,160	34
H1-H2	Sample 2	unknown	0,588	0,513	115
A3-A4	Sample 3	unknown	1,414	1,342	332

### Quality control

Each kit contains FITkit® Hev b 1 Control, which should give results within the specified range given in a separate certificate of analysis enclosed in the kit.



## 9. Expected values

Results obtained from 13 different glove brands extracted to PBS (1 g glove/5 mL PBS) in 2 hours at room temperature are given below in Table 1.

**Table 1. Hev b 1 content of different glove brands extracted in PBS**

Extract	Type	Glove Material	Hev b 1 µg/L
1	Examination	Latex (powdered)	17
2	Examination	Latex (powder free)	<10
3	Examination	Latex (powder free)	<10
4	Examination	Latex (powder free)	50
5	Examination	Latex (powdered)	<10
6	Surgical	Latex (powder free)	<10
7	Surgical	Latex (powder free)	<10
8	Surgical	Synthetic (powdered)	<10
9	Examination	Nitrile (protein free)	<10
10	Examination	Vinyl	<10
11	Surgical	Latex (powdered)	<10
12	Surgical	Latex (powdered)	23
13	Surgical	Latex (powder free)	11



## 10. Performance characteristics

### 10.1 Detection limit

The detection limit of Hev b 1 test was defined by the minimum Hev b 1 concentration deviating by 2 SD from that of the Calibrator A. The test was performed by using 16 replicate determinations of the Calibrator A and Calibrator B. On the basis of this test the detection limit of Hev b 1 assay is 1.2 µg/L.

### 10.2 Precision

Repeatability (intra-assay variation) and reproducibility (inter-assay variation) were determined by analysing three samples containing a low, medium and high concentration of Hev b 1. The results are given in Table 2 and 3.

**Table 2. Repeatability**

Sample	Number of replicates	Mean (µg/L)	SD (µg/L)	CV%
1	16	10	0.6	5.6
2	16	130	6.0	4.6
3	16	321	9.0	2.8

**Table 3. Reproducibility**

Sample	Number of assays	Mean (µg/L)	SD (µg/L)	CV%
1	5	22	1.5	6.8
2	5	34	1.9	5.6
3	5	231	10.5	4.6



### 10.3 Recovery

50, 200 and 1000 µg/L concentrations of purified Hev b 1 calibrator were added to equal volumes of three samples containing a low (34 µg/L), medium (121 µg/L) and high (327 µg/L) concentration of Hev b 1. Determination of Hev b 1 was done using unspiked samples and samples spiked with Hev b 1 calibrators. The theoretical concentration and the recovered concentrations were calculated. The results are shown in Table 4.

**Table 4. Recovery**

Sample	Added conc. (µg/L)	Expected conc. (µg/L)	Obtained conc. (µg/L)	Recovery %
Low	0		34	100
	50	42	49	117
	200	117	102	87
	1000	517	639	124
Medium	0		121	100
	50	85	95	112
	200	160	179	112
	1000	560	667	119
High	0		327	100
	50	188	199	106
	200	263	243	92
	1000	663	691	104

### 10.4 Linearity (dilution test)

Three samples containing 31, 128 and 296 µg/L of Hev b 1, were diluted with PBS 1:2, 1:5, 1:10 and 1:20, if appropriate. The concentration of each diluted and original sample was measured. The results given as the percentage of the original concentration corrected with the dilution factors are given in Table 5.

**Table 5. Linearity**

Sample	Dilution	Conc. (µg/L)	%
1	undiluted	31	100
	1:2	32	103
2	undiluted	128	100
	1:2	131	102
	1:5	131	100
	1:10	134	102
3	undiluted	296	100
	1:2	283	96
	1:5	309	104
	1:10	347	117
	1:20	392	132

### 10.5 Specificity

#### Cross-reactions

Cross-reactions with Hev b 6.02, Hev b 3, Hev b 5 and Hev b 7 were tested using concentrations on a weight-to-weight basis. No cross reactions were detected, thus the cross reactivities are less than 0.01 %, 0.05 %, 0.02 % and 0.01 %, respectively. The results are given in Table 6.

**Table 6. Cross-reactions**

Substance	Concentration tested	Cross-reaction (w/w)
Hev b 6.02	100 000 µg/L	< 0.01 %
Hev b 3	20 000 µg/L	< 0.05 %
Hev b 5	50 000 µg/L	< 0.02 %
Hev b 7	86 000 µg/L	< 0.01 %

## Interfering substances

Several accelerators, antioxidants and detergents used in the rubber industry were tested using the interfering substance in PBS and in the solution containing Hev b 1. The test was performed in the presence and absence of each substance.

Substances tested: 0.1 % and 0.02 % solutions of pure ZDEC, ZDBC, SDBC, AS100, Arbostab Z, ZMBT, P25, BKF, Ralox LC, MB2, Setsit 104 and 0.1 % and 0.02 % (v/v) solutions of ZDEC, ZDBC, ZMBT, MBT, TMTD, DPTT, ZnO, Sulfur, TiO<sub>2</sub>, Wingstay made from dispersions thereof. Inhibition of binding varied 0 – 45 %.

Solutions containing 0.005 % of Triton X114, Surfynol TG, Surfynol DF37, Foamaster VL, Sodium Caprylate, Cellosize, Igepal CA630 and Alginate N40 were tested in the same way, inhibition of binding was less than 18 %. Darvan #1 in the 0.0005 % solution inhibited the binding of Hev b 1 by 25 %.

No substance alone gave any background in the assay.



## 11. Additional information

Current FITkit® Hev b 1 is one of four FITkits available. There are specific kits to measure Hev b 3, Hev b 5 and Hev b 6.02.

Additional information about FITkits and for kit users (troubleshooting, frequently asked questions, microwell plate layout print version etc.) is available on the web page [www.icosagen.com](http://www.icosagen.com).

## 12. Literature

Yeang HY, Cheong KF, Sunderasan E, Hamzah S, Chew NP, Hamid S, Hamilton RG, Cardoso J. The 14.6 kD rubber elongation factor (Hev b 1) and 24 kD (Hev b 3) rubber particle proteins are recognized by IgE from patients with spina bifida and latex allergy. *J Allergy Clin Immunol* 1996; 98: 628-639

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