



本プロトコールは参考用の資料になります。商品ご購入の際は必ず商品に添付されている資料をご参照ください。

# ***Progesterone ELISA Kit Instructions***

***Please read all instructions carefully before beginning this assay***

PRODUCT #402310  
For Research Use Only

**\*\*\*Store kit at 4°C at all times\*\*\***

**Do not freeze kit components**

## **DESCRIPTION**

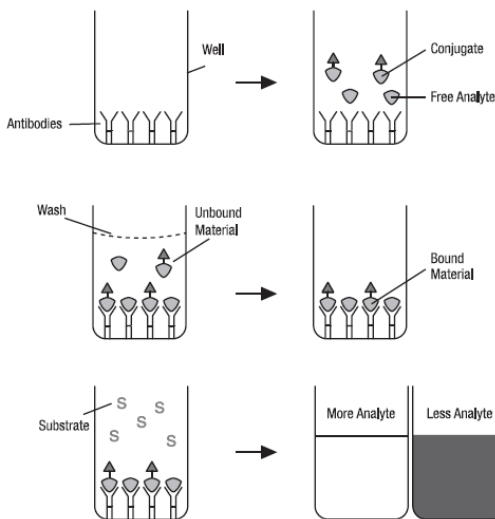
Progesterone is a C-21 steroid secreted by the corpus luteum and the placenta. It is an important intermediate in steroid biosynthesis in all tissues that secrete steroid hormones. Progesterone is responsible for the progestational changes in the endometrium and cyclic changes in the cervix and vagina. The plasma progesterone level is useful in investigating the luteal function and monitoring ovulation.

## **PRINCIPLE OF ASSAY**

This is an ELISA (Enzyme-Linked Immunosorbent Assay) for the quantitative analysis of Progesterone levels in biological fluid. This test kit operates on the basis of competition between the enzyme conjugate and the Progesterone in the sample for a limited number of binding sites on the antibody coated plate.

The sample or standard solution is first added to the microplate. Next, the diluted enzyme conjugate is added and the mixture is shaken and incubated at room temperature for one hour. During the incubation, competition for binding sites is taking place. The plate is then washed removing all the unbound material. The bound enzyme conjugate is detected by the addition of substrate which generates an optimal color after 30 minutes. Quantitative test results may be obtained by measuring and comparing the absorbance reading of the wells of the samples against the standards with a microplate reader at 650 nm. The extent of color development is inversely proportional to the amount of Progesterone in the sample or standard. For example, the absence of Progesterone in the sample will result in a bright blue color, whereas the presence of Progesterone will result in decreased or no color development.

## PRINCIPLE OF ASSAY (continued)



## MATERIALS PROVIDED

1. EIA BUFFER: 30 mL. Provided to dilute enzyme conjugate and Progesterone standards.
2. WASH BUFFER (10X): 20 mL. Dilute 10-fold with deionized water. Diluted wash buffer is used to wash all unbound enzyme conjugate, samples and standards from the plate after the one hour incubation.
3. K-BLUE SUBSTRATE: 20 mL. Stabilized 3,3',5,5' Tetramethylbenzidine (TMB) plus Hydrogen Peroxide ( $H_2O_2$ ) in a single bottle. It is used to develop the color in the wells after they have been washed. Keep substrate refrigerated. LIGHT SENSITIVE
4. EXTRACTION BUFFER (5X): 30 mL. Diluted 5-fold with deionized water. This buffer is used for diluting extracted and non-extracted samples.
5. PROGESTERONE ENZYME CONJUGATE: 150  $\mu$ L. Progesterone horseradish peroxidase concentrate. Blue capped vial.
6. PROGESTERONE STANDARD: 100  $\mu$ L. Progesterone standard at the concentration of 1  $\mu$ g/mL. Green capped vial.
7. PROGESTERONE ANTIBODY-COATED MICROPLATE: A 96 well MaxiSorp™ Nunc microplate with anti-Progesterone rabbit antibody precoated on each well. The plate is ready for use as is. DO NOT WASH!

## MATERIALS NEEDED BUT NOT PROVIDED

1. 300 mL deionized water for diluting wash buffer and extraction buffer.
2. Precision pipettes that range from 10  $\mu$ L-1000  $\mu$ L and disposable tips.

NOTE: If all or several strips are to be used at one time, it is suggested that a multichannel pipette be used.

3. Clean test tubes used to dilute the standards and conjugate.
4. Graduated cylinders to dilute and mix wash buffer and extraction buffer.
5. Microplate reader with 650 nm filter.
6. Plate cover or plastic film to cover plate during incubation.

## OPTIONAL MATERIALS:

7. 1N HCl or Neogen Corporation's Red Stop Solution.
8. Microplate shaker.

If performing an extraction on samples, the following will be required:

9. Petroleum ether
10. Nitrogen gas
11. Vortex

## **WARNINGS AND PRECAUTIONS**

1. DO NOT use components beyond expiration date.
2. DO NOT mix any reagents or components of this kit with any reagents or components of any other kit. This kit is designed to work properly as provided.
3. DO NOT pipette reagents by mouth.
4. Always pour substrate out of the bottle into a clean test tube - DO NOT pipette out of the bottle (if your tip is unclear you could contaminate your substrate).
5. All specimens should be considered potentially infectious. Exercise proper handling precautions.
6. DO NOT smoke, eat or drink in areas where specimens or reagents are being handled.
7. Use aseptic technique when opening and removing reagents from vials and bottles.
8. Keep plate covered except when adding reagents, washing or reading.
9. Kit components should be refrigerated at all times when not in use.

## **PROCEDURAL NOTES**

1. It is not necessary to allow reagents to warm to room temperature before use.
2. Desiccant bag must remain in foil pouch with unused strips. Keep pouch sealed when not in use to maintain a dry environment. Try to remove excess air before sealing.
3. Always use different pipette tips for the buffer, enzyme conjugate, standards and samples.
4. Before pipetting a reagent, rinse the pipette tip three times with that reagent (i.e. fill the tip with the desired amount of reagent and dispense back into the same vial - repeat 2 times). Now the tip is properly rinsed and ready to dispense the reagent into your well or test tube.
5. When pipetting into the wells, DO NOT allow the pipette tip to touch the inside of the well, or any of the reagents already in the well - this can cause cross contamination.
6. Standards and samples should be assayed in duplicate.
7. To quantitate, always run a standard curve when testing samples. If testing a sample that is not extracted, dilute standards in the same type of medium being tested, which is known to be negative.
8. Gently mix specimens and reagents before use. Avoid vigorous agitation.
9. When using only partial amounts of a kit, it is recommended to transfer the appropriate volume of each reagent to a clean vessel for repeated dispensing. This will reduce reagent contamination by repeated sampling from the original container.
10. The enzyme conjugate is most stable in its concentrated form. Dilute only the volume necessary for the amount of strips currently being used.
11. Before taking an absorbance reading wipe the outside bottom of the wells with a lint-free wiper to remove dust and fingerprints.
12. Before opening the enzyme conjugate and standard vial, tap vial in an upright position to remove any liquid in the cap.

## **SAMPLE PREPARATION**

This assay is non-species specific. Usually, urine and tissue culture supernatant can be assayed directly by diluting them with the diluted extraction buffer. Plasma and most other mediums will need to be extracted.

## EXTRACTION OF PROGESTERONE

1. Pipette 100  $\mu\text{L}$  of plasma into a glass tube (10x75 mm) and add 1 mL of petroleum ether.
2. Vortex the tube for 30 seconds and then allow the phases to separate.
3. Transfer the organic phase into a clean glass tube and evaporate the solvent with a stream of  $\text{N}_2$ .
4. Dissolve the residue in 500  $\mu\text{L}$  of diluted extraction buffer.
5. Vortex and assay 50  $\mu\text{L}$  in duplicates.
6. The values obtained are multiplied by 5 to give final ng/mL concentrations.
7. Assay the following volume for plasma concentrations:  
Male: 50  $\mu\text{L}$  in duplicate wells  
Female: 10  $\mu\text{L}$  duplicate wells (Add 40  $\mu\text{L}$  of diluted extraction buffer in these duplicate wells.)
8. If the concentration is higher than the high range of the standard curve, the above extract should be further diluted and reassayed.

NOTE: Extraction buffer must be diluted 5-fold with deionized water before use. Any precipitant present must be brought into solution before dilution.

## TEST PROCEDURES

1. Prepare standards as follows:

Standard	Preparation
A	stock solution 1 $\mu\text{g/mL}$ (Provided in green capped vial.)
B	take 20 $\mu\text{L}$ of A, add to 980 $\mu\text{L}$ of EIA buffer and mix=20 ng/mL
C	take 100 $\mu\text{L}$ of B, add to 900 $\mu\text{L}$ of EIA buffer and mix=2 ng/mL

Continue standard preparation following Scheme I.

### SCHEME I

Standards	ng/mL	EIA buffer ( $\mu\text{L}$ added)	A standard $\mu\text{L}$	B standard $\mu\text{L}$	C standard $\mu\text{L}$
$S_0$	0	as is	-	-	-
$S_1$	0.4	400	-	-	100
$S_2$	1	250	-	-	250
$S_3$	2	-	-	-	as is
$S_4$	4	400	-	100	-
$S_5$	10	250	-	250	-
$S_6$	20	-	-	as is	-
$S_7$	40	480	20	-	-

2. Determine the number of wells to be used.
3. Dilute the Progesterone enzyme conjugate. Add 1  $\mu\text{L}$  of enzyme conjugate into 50  $\mu\text{L}$  total volume of EIA buffer for each well assayed. For the whole plate, add 110  $\mu\text{L}$  of the enzyme conjugate into 5.5 mL total volume of EIA buffer. Mix the solution thoroughly.
4. Add 50  $\mu\text{L}$  of standards (S) or unknown (U) (some samples may require diluting) to the appropriate wells in duplicate. See Scheme II for suggested template design.
5. Add 50  $\mu\text{L}$  of the diluted enzyme conjugate to each well. Use 8-channel pipette or 12-channel pipette for rapid addition.
6. Mix by shaking plate gently. A microplate shaker may be used.

- Cover plate with plastic film or plate cover and incubate at room temperature for one hour. NOTE: Keep plate away from drafts and temperature fluctuations.
- Dilute concentrated wash buffer with deionized water (i.e. 20 mL of wash buffer plus 180 mL of deionized water). Mix thoroughly.
- After incubation, dump out the contents of the plate. Tap out contents thoroughly on a clean lint-free towel.
- Wash each well with 300  $\mu$ L of the diluted wash buffer. Repeat for a total of three washings. An automated plate washer can be used, however, increase number of wash cycles from three to five.
- Add 150  $\mu$ L of substrate to each well. Use multichannel pipette for best results. Mix by shaking plate gently.
- Incubate at room temperature for 30 minutes.
- Gently shake plate before taking a reading to ensure uniform color throughout each well.
- Plate is read in a microplate reader at 650 nm. If a dual wavelength is used, set  $W_1$  at 650 nm and  $W_2$  at 490 nm.
- If accounting for substrate background, use 2 to 8 wells as blanks with only substrate in the wells (150  $\mu$ L/well). Subtract the average of these absorbance values from the absorbance values of the wells being assayed.

NOTE: Some microplate readers can be programmed to do these subtractions automatically when reading the plate. Consult your instrument manual.

## OPTIONAL TEST PROCEDURES

- Add 50-100  $\mu$ L of 1N HCl or Neogen's Red Stop Solution to each well to stop enzyme reaction.
- Read plate at 450 nm, if 1N HCl solution was used. Read plate at 650 nm, if Neogen's Red Stop Solution was used.
- Plot the standard curve and estimate the concentrations of the samples from the curve. See "CALCULATIONS."

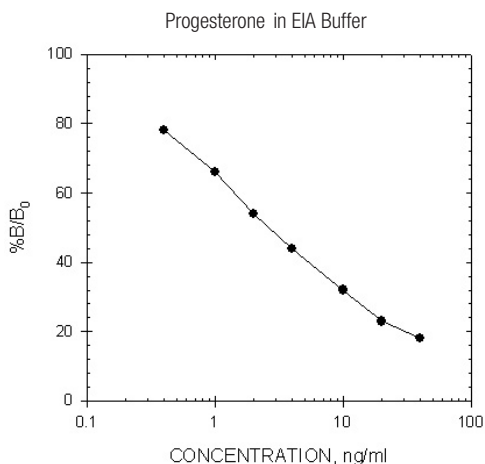
NOTE: Absorbance readings will approximately double when stopped with acid. If absorbance readings are too high for measuring with your microplate reader, decrease the substrate incubation approximately 10 minutes but no more than 15 minutes.

## SCHEME II

	1	2	3	4	5	6	7	8	9	10	11	12
A	$S_0$	$S_0$	$U_1$	$U_1$	$U_9$	$U_9$	$U_{17}$	$U_{17}$	$U_{25}$	$U_{25}$	$U_{33}$	$U_{33}$
B	$S_1$	$S_1$	$U_2$	$U_2$	$U_{10}$	$U_{10}$	$U_{18}$	$U_{18}$	$U_{26}$	$U_{26}$	$U_{34}$	$U_{34}$
C	$S_2$	$S_2$	$U_3$	$U_3$	$U_{11}$	$U_{11}$	$U_{19}$	$U_{19}$	$U_{27}$	$U_{27}$	$U_{35}$	$U_{35}$
D	$S_3$	$S_3$	$U_4$	$U_4$	$U_{12}$	$U_{12}$	$U_{20}$	$U_{20}$	$U_{28}$	$U_{28}$	$U_{36}$	$U_{36}$
E	$S_4$	$S_4$	$U_5$	$U_5$	$U_{13}$	$U_{13}$	$U_{21}$	$U_{21}$	$U_{29}$	$U_{29}$	$U_{37}$	$U_{37}$
F	$S_5$	$S_5$	$U_6$	$U_6$	$U_{14}$	$U_{14}$	$U_{22}$	$U_{22}$	$U_{30}$	$U_{30}$	$U_{38}$	$U_{38}$
G	$S_6$	$S_6$	$U_7$	$U_7$	$U_{15}$	$U_{15}$	$U_{23}$	$U_{23}$	$U_{31}$	$U_{31}$	$U_{39}$	$U_{39}$
H	$S_7$	$S_7$	$U_8$	$U_8$	$U_{16}$	$U_{16}$	$U_{24}$	$U_{24}$	$U_{32}$	$U_{32}$	$U_{40}$	$U_{40}$

- After the substrate background has been subtracted from all absorbance values, average all of your duplicate well absorbance values.
- The average of your two  $S_0$  values is now your  $B_0$  value. ( $S_1$  now becomes  $B_1$ , etc.)
- Next, find the percent of maximal binding ( $\%B/B_0$  value). To do this, divide the averages of each standard absorbance value (now known as  $B_1$  through  $B_7$ ) by the  $B_0$  absorbance value and multiply by 100 to achieve percentages.
- Graph your standard curve by plotting the  $\%B/B_0$  for each standard concentration on the ordinate (y) axis against concentration on the abscissa (x) axis. Draw a curve by using a curve-fitting routine (i.e. 4-parameter or linear regression).
- Divide the averages of each sample absorbance value by the  $B_0$  value and multiply by 100 to achieve percentages.
- Using the standard curve, the concentration of each sample can be determined by comparing the  $\%B/B_0$  of each sample to the corresponding concentration of Progesterone standard.
- If the samples were diluted, the concentration determined from the standard curve must be multiplied by the dilution factor.

## TYPICAL STANDARD CURVE



## TYPICAL DATA

NOTE: "Typical data" is a representation. Variances in data will occur. Optical density readings may fluctuate during the shelf life of the kit, but the %B/B<sub>0</sub> should remain comparable. Measuring wavelength: 650 nm

Standard	Standard Concentration (ng/mL)	Optical Density (Absorbance Value)	%B/B <sub>0</sub>
S <sub>0</sub> (B <sub>0</sub> )	0	1.200	100
S <sub>1</sub> (B <sub>1</sub> )	0.4	0.940	78
S <sub>2</sub> (B <sub>2</sub> )	1	0.788	66
S <sub>3</sub> (B <sub>3</sub> )	2	0.647	54
S <sub>4</sub> (B <sub>4</sub> )	4	0.526	44
S <sub>5</sub> (B <sub>5</sub> )	10	0.378	32
S <sub>6</sub> (B <sub>6</sub> )	20	0.278	23
S <sub>7</sub> (B <sub>7</sub> )	40	0.214	18

## CROSS REACTIVITY

PROGESTERONE.....	100.0%
DEOXYCORTICOSTERONE.....	2.5%
CORTICOSTERONE.....	2.0%
PREGNENOLONE.....	2.0%
ANDROSTENEDIONE.....	1.0%
17-HYDROXYPROGESTERONE.....	0.40%
TESTOSTERONE.....	0.29%
CORTISOL.....	0.20%
CORTISONE.....	0.20%
DEHYDROEPIANDROSTERONE.....	0.20%
ESTRADIOL.....	0.20%
ESTRIOL.....	0.20%
ESTRONE.....	0.20%

## COPYRIGHT

All rights reserved worldwide. No part of this publication may be reproduced, transmitted, transcribed, or stored in any information-retrieval system, or translated into any human or computer language in any form or by any means (manual, electronic, mechanical, magnetic, optical, chemical, or otherwise) without expressed written permission.

## WARRANTY

Neogen Corporation makes no warranty of any kind, either expressed or implied, except that the material from which its products are made are of standard quality. If any materials are defective, Neogen Corporation will provide a replacement product. Buyer assumes all risk and liability resulting from the use of this product and any of the predictive models. There is no warranty of merchantability of this product, or of the fitness of the product for any purpose. Neogen Corporation shall not be liable for any damages, including special or consequential damage, or expense arising directly or indirectly from the use of this product.

## TECHNICAL ASSISTANCE

Technical assistance is available Monday-Friday between 8:00 a.m. and 6:00 p.m. EST.



944 Nandino Blvd • Lexington KY 40511-1205 USA  
859/254-1221 or 800/477-8201 USA/CANADA  
Fax: 859/255-5532 • Email: [inform@neogen.com](mailto:inform@neogen.com)