

PRODUCT PROFILE AND INSTRUCTIONS
LYOPHILIZED STANDARDS**INTENDED USE:**

The Cortisol ELISA test is an immunoassay designed for the quantitative determination of cortisol in serum/plasma. The test is intended for professional use as an aid in the diagnosis and monitoring of physiological/pathological conditions related to serum/plasma cortisol in Swine and related species.

PRINCIPLES OF TEST

The Cortisol Quantitative Test is based on a widely used immunoassay technique. A sample (serum/ plasma) containing an unknown amount of cortisol to be assayed (unlabeled antigen) is added to a standard amount of a labeled derivative of the same substance (labeled antigen). The labeled and unlabeled antigens are then allowed to compete for high affinity binding sites on a limited number of antibodies coated on to the plate. After washing away the free antigen, the amount of labeled antigen in the sample is reversibly proportional to the concentration of the unlabeled antigen. The actual concentrations in unknown samples are obtained by means of a standard curve based on known concentrations of unlabeled antigen analyzed in parallel with the unknowns. In this kit an enzyme label is used. The bio-specific reaction takes place during 1 hour incubation. After washing away, substrate solution is added and the enzyme allowed to react for a fixed time before the reaction is terminated. Absorbencies are measured at 450 nm using ELISA plate reader. A standard curve is produced using values from 6 standards from which absorbency values for blank tubes have been subtracted. Results for unknown may be read directly from this standard curve using either manual calculation or by a suitable computer program.

This kit is suitable for the direct measurement of cortisol in serum/plasma samples.

Note: The cortisol levels should be established in your laboratory using your own set of samples and standards and good laboratory practice should be employed where ever is applicable.

Materials Provided

1. Microtiter wells coated with cortisol specific antibody.
2. HRP Conjugate, 12mL
3. Cortisol Standards, Lyophilized. (Contains 0, 1.0, 2.5, 5.0,10,50, 200 ng/mL), dilute to 0.6 ml/vial with Standard/Sample Diluent.
4. TMB Color Reagent, 12 mL
5. Stop Solution (2N HCl), 6 mL
6. 20x Wash Buffer, 20mL.
7. Standard/Sample diluent, 20mL

Materials Required, but Not Provided

1. Semiautomatic pipettes: 20ul and 200ul
2. Disposable pipette tips
3. Microtiter plate shaker
4. Microtiter well reader.
5. Plate washer
6. Absorbant paper
7. 37 C incubator
8. Parafilm to cover plate
9. Distilled water

PRECAUTIONS

1. CAUTION: This kit contains reagents manufactured from serum/plasma components. The source materials have been tested by immunoassay for hepatitis B surface antigen and antibodies to HIV virus and found to be negative. Nevertheless, all blood products and samples should be considered potentially infectious and handling should be in accordance with the procedures defined by an appropriate biohazard safety guideline or regulations in your labs or local and state.

2. The contents of this kit, and their residues, must not come into contact ruminating animals or swine.
3. Avoid contact with the Stopping Reagent. It may cause skin irritation and burns.
4. Do not use reagents after expiration date.
5. Do not mix or use components from the kits with different lot numbers.
6. Replace caps on reagents immediately. Do not switch caps.
7. Reagents contain sodium azide (NaN₃) as a preservative. On disposal, flush with a large volume of water to prevent azide build-up.
8. Do not pipette reagents by mouth.
9. Do not use reagents from other kits or mix with other manufactured test kits.

STORAGE CONDITIONS

1. Store the kit at 4-8 C upon receipt and when it is not in use.
2. Keep microtiter wells in a sealed bag with desiccants to minimize exposure to damp air.
3. After every use place the caps tightly. Store reagents at 4-8°C and standards at -20°C.

INSTRUMENTATION

A microtiter well reader with bandwidth of 10 nm or less and an optical density range of 0 to 2 OD or greater at 450 nm wavelength is acceptable for use in absorbency measurement.

Sample Collection and Reagent Preparation.

1. This kit is suitable for use with serum or plasma samples. The use of grossly hemolytic or lipemic samples should not be used may affect results. Samples with bilirubin may also interfere with the assay. A venous blood sample (enough to produce about 0.5 ml serum) is collected aseptically.



2. Lyophilized standards should reconstitute with 0.6ml standard diluent (and be kept frozen at -20C if not used immediately). Make sure all the tube contents must be dissolved completely before use in the assay.
3. Dilute wash buffer, desired amount with distilled water (1part with 19 parts). The buffer is stable for 1-3 months, if stored at 4-8C.
4. Highly concentrated samples should be diluted with sample diluent (eg. 1:5, or 1:10), to bring on to a readable range on the curve. The samples diluted can be stored at -20°C for further use.

ASSAY PROCEDURE

1. Pipette 25ul of standards.
2. Add 25ul of test samples into appropriate wells.
3. Add 100ul of Cortisol Enzyme Conjugate Solution to each well (except those set aside for blanks).
4. Incubate for 1 hour at 37C.
5. Terminate the reaction and wash the plate 4-5 times with Wash Solution (250-300ul) per well. Invert plate, tap firmly against absorbent paper to remove any residual moisture.
6. Add 100 ul of TMB color reagent into each well (including the blanks). Remember for pipetting order.
7. Incubate the plate for 20 minutes without shaking.
8. Stop reaction by adding 50ul of Stopping Solution (a drop) to each well in the same sequence that the Substrate Solution was added. Gently mix for 1-2 minutes.
9. Read the absorbency at 450 nm with a microwell reader.

NOTE: The substrate incubation should be carried out at room temperature (within the temperature range 25-28C). For temperature outside this range, the duration of the incubation should be adjusted by approximately 1 minute/1C.

CALCULATION OF RESULTS

1. Calculate the mean absorbance values (A) for each set of reference standards, controls, samples and blanks.
2. Subtract the value for blanks from those for standards, control and unknown samples.
3. Calculate the B/B% values by dividing each value by the value for the zero-standard.
4. For the standards, plot a graph on semi-log graph paper with B/BO% values on the ordinate and the Cortisol concentrations (ng/mL) on the abscissa.
5. Using the graph read off the Cortisol concentrations for the unknown samples.
6. You may use any commercial assay software to analyze the data.

EXPECTED VALUES AND SENSITIVITY

It is recommended that each laboratory should establish values to reflect differences specific to experimental conditions. The minimum detectable concentration of cortisol by this assay is estimated to be 0.2ng/ml.

Limitations & Warranty

The present Swine cortisol ELISA is designed for helping the scientist to analyze test samples from Swine species only. There are no warranties, expressed, implied or otherwise indicated, which extend beyond this description of this product. Endocrine Technologies, Inc. is not liable for property or laboratory damage, personal injury, or test samples loss, or economic loss caused by this product. Warranty is limited to replacement of similar ELISA Kit or reagents damaged during shipment or leaking solutions or any other issues within 30 days, with written explanation, with sharing the entire data including raw printouts from the reader and return of the ELISA product. The analyst should establish the standard curve and a small number of samples before proceeding to analyze a large number of samples. Endocrine will not replace or refund, only if deemed necessary on scientific merits within 30 days only.

REFERENCES

1. BONDY PK. The adrenal cortex, in Randy PT, Rosenberg LE., Metabolic control and disease (8 ed) 1980, WB Sanders, Philadelphia, p1427-1499.
2. Lambert A Clinical Endocrinology 1974, Springer-Verlag New York, p299-305
3. Veosei P, Glucocorticoids, Cortisol, corticosterone, Compound S in Jaffe BM, Behrman HR 1974 in Methods in Radioimmuno assay p393-411.
4. Spark R 1971, Simplified assessment of pituitary-adrenal reserve Annals of internal Med. 75 p75-717
2. Wise T, Zanella EL, Lunstra DD, Ford JJ. Relationships of gonadotropins, testosterone, and cortisol in response to GnRH and GnRH antagonist in boars selected for high and low follicle-stimulating hormone levels. J Anim Sci 2000 Jun;78(6):1577-1590
6. Mariscal DV, Bergfeld EG, Cupp AS, Kojima FN, Fike KE, Sanchez T, Wehrman ME, Johnson RK, Kittok RJ, Ford JJ, Kinder JE. Concentrations of gonadotropins, estradiol and progesterone in sows selected on an index of ovulation rate and embryo survival. Anim Reprod Sci 1998 Dec 1;54(1):31-43
4. Perremans S, Randall JM, Rombouts G, Decuypere E, Geers R. Effect of whole-body vibration in the vertical axis on cortisol and adrenocorticotrophic hormone levels in piglets. J Anim Sci 2001 Apr;79(4):975-981
5. Schonreiter S, Huber H, Lohmuller V, Zanella AJ, Unshelm J, Henke J, Erhardt W. Tierarztl Prax Ausg G Grosstiere Salivary cortisol as a stress parameter in piglets Nutztiere 1999 May;27(3):175-179

Revised 0217

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