



## REAGENTS

**96 well polystyrene microplate (12 strip of 8 well) coated with Tat protein and blocked with BSA 1%**

**Plate Cover :** 1 adhesive plate sealers

**Assay diluent:** (Buffer A) 50 mL ready to use, with preservative

**Wash buffer concentrate:** (Buffer B) 125 mL 10x to dilute to 1250 final volume with distilled water, with preservative.

**Conjugated Antibody:** Goat Anti rabbit IgG-HRP conjugated 20µl

**Color buffer:** (Buffer C) 12.5 mL with preservative

**1 tablets of ABTS** (Buffer C<sub>1</sub>) 12.5µl

## STORAGE

Maintain the kit at 2-8°C. Do not use kit past its expiration date.

## OTHER SUPPLIES REQUIRED

- Microplate reader capable of measuring absorbance at 405 nm.
- Pipettes and pipette tips.
- Deionized or distilled water
- Multi-channel pipette, squirt bottle, manifold dispenser, or automated microplate washer
- 2000 mL graduated cylinder for preparation of Wash Buffer
- vortex mixer
- Glass Vials

## PRECAUTIONS

The buffer C1 provided with this kit is H<sub>2</sub>O<sub>2</sub> solution 30% m/m (110 volumes) causes burns: after contact with skin, wash immediately with plenty of water. Wear suitable protective clothing and eye/face protection.

The antibody used in this assay as no testing can offer complete assurance of freedom from infectious agents.

## SAMPLE COLLECTION AND STORAGE

**Serum:** Use pyrogen/endotoxin free collecting tubes. Serum should be removed rapidly and carefully from the red cells after clotting. For that reason centrifuge, after clotting, at approximately 1000 x g for 10 min and remove serum.

**Plasma samples** should be collected in heparin or EDTA and centrifuged at 1000 x g for 10 min; plasma should be removed rapidly and carefully from the red cells after centrifugation.

**Storage:** If not analyzed shortly after collection, freeze samples in small aliquots (25-50µl) and store them at -80°C. Avoid freeze-thaw cycles.

**Recommendation:** Do not thaw by heating at 37°C or 56°C. Thaw at room temperature and make sure that sample is completely thawed and homogeneous before assaying.

## REAGENT PREPARATION

Bring all reagents to room temperature prior to use.

**96 wells microtiter plates:** balance the plate at room temperature for 1 h before use.

**Assay diluent:** ready to use. Utilize this buffer for sera or plasma dilutions in a clean glass vial.

**Wash Buffer concentrate:** If crystals have formed in the concentrate, warm to room temperature and mix gently until the crystals have completely dissolved. Dilute 125 mL of Wash Buffer ( Buffer B) concentrate into deionized or distilled water to prepare 1250 mL of Wash Buffer.

**Conjugated antibody:** Prepare immediately before use. Dilute 1:1000 in buffer A, in a clean glass vial according to the number of wells to be used (100µl / well). Do not keep this dilution for further experiments.

**Color buffer:** reconstitute immediately before use the ABTS tablet in 12.5 mL of buffer C and mix with 12.5 µl of buffer C<sub>1</sub>. The Color buffer is light sensitive. Avoid prolonged exposure to light.

## ASSAY METHOD

Bring all reagents and samples to room temperature before use. It's recommended that all sample and standards be assayed in duplicate.

- 1 Take away the sealing from the plates
- 2 Dispense the serum at appropriate dilution in buffer A (100 µl / well)
- 3 Maintain the plates 90 min. at 37°C: cover the plates.
- 4 Wash the plates five times (in diluted buffer B)
- 5 Dispense the Goat anti mouse IgG HRP-conjugated diluted 1:1000 in buffer A (100 µl / well)
- 6 Maintain the plates for 60 min. at 37°C: cover the plates.
- 7 Wash the plates five times ( in diluted buffer B)
- 8 Dispense the reconstituted color buffer (100 µl/well)
- 9 Cover the plates. Await the development of the color at room temperature: usually it takes from a minimum of 20 min. to a maximum of 45 min.
- 10 Read the abs at 405 nm using a microplate reader

## QUALITY CONTROL

Each testing laboratory should establish a quality control program to monitor the performance of the Rabbit Anti-TAT IgG Immunoassay.

## ASSAY PROCEDURE SUMMARY

- Dispense the serum at appropriate dilutions 100 µl /well  
Maintain the plates at 37°C for 90 min.
- Aspirate and wash five times.
- Add 100µl per well of conjugated antibody  
Maintain the plates at 37°C for 60 min
- Aspirate and wash five times
- Add 100 µl / well of Color Buffer  
Incubate 20-45 min. RT, Read at 405 nm

## REFERENCES

- Arya SK, Guo C, Josephs SF, Wong-Staal F. “The trans-activator gene of human T- Lymphotropic virus type III (HTLV-III)” *Science*. 1985 Jul 5;229(4708):69-73
- Reiss P, Lange JM, de Ronde A, de Wolf F, Dekker J, Deboucq C, Goudsmit J: “Speed of progression to AIDS and degree of antibody response to accessory gene products of HIV-1” *J Med Virol*. 1990 Mar;30(3):163-8
- Ensoli B, Buonaguro L, Barillari G, Fiorelli V, Gendelman R, Morgan RA, Wingfield P, Gallo RC: “Release, uptake, and effects of extracellular human immunodeficiency virus type 1 Tat protein on cell growth and viral *trans* activation” *J Virol*. 1993 Jan;67(1):277-87
- Chang HK, Gallo RC, Ensoli B “Regulation of cellular gene expression and function by the human immunodeficiency virus type 1 Tat protein” *J Biomed Sci*. 1995 Aug;2(3):189-202
- Re MC, Furlini G, Vignoli M, Ramazzotti E, Roderigo G, De Rosa V, Zauli G, Lolli S, Capitani S, La Placa M: “Effect of antibody to HIV-1 Tat protein on viral replication in vitro and progression of HIV-1 disease in vivo” *J Acquir Immune Defic Syndr Hum Retrovirol*. 1995 Dec 1;10(4):408-16
- Fauci AS: “An HIV vaccine: breaking the paradigms” *Proc Assoc Am Physicians*. 1996 Jan;108(1):6-13. Review.
- Borrow P, Lewicki H, Wei X, Horwitz MS, Pfeffer N, Meyers H, Nelson JA, Gairin JE, Hahn BH, Oldstone MB, Shaw GM: “Antiviral pressure exerted by HIV-1-specific cytotoxic T lymphocytes (CTLs) during primary infection demonstrated by rapid selection of CTL escape virus” *Nat Med*. 1997 Feb;3(2):205-11
- Kim DT, Mitchell DJ, Brockstedt DG, Fong L, Nolan GP, Fathman CG, Engleman EG, Rothbard JB: “Introduction of soluble proteins into the MHC class I pathway by conjugation to an HIV tat peptide” *J Immunol*. 1997 Aug 15;159(4):1666-8
- Chang HC, Samaniego F, Nair BC, Buonaguro L, Ensoli B: “HIV-1 Tat protein exits from cells via a leaderless secretory pathway and binds to extracellular matrix-associated heparan sulfate proteoglycans through its basic region” *AIDS*. 1997 Oct;11(12):1421-31.
- Zagury JF, Sill A, Blattner W, Lachgar A, Le Buanec H, Richardson M, Rappaport J, Hendel H, Bizzini B, Gringeri A, Carcagno M, Criscuolo M, Burny A, Gallo RC, Zagury D: “Antibodies to the HIV-1 Tat protein correlated with nonprogression to AIDS. A rationale for the use of Tat toxoid as an HIV-1 vaccine” *J Hum Virol*. 1998 May-Jun;1(4):282-92
- Re MC, Vignoli M, Furlini G, Gibellini D, Colangeli V, Vitone F, La Placa M “Antibodies against full-length Tat protein and some low-molecular- weight Tat-peptides correlate with low or undetectable viral load in HIV-1 seropositive patients” *J Clin Virol*. 2001 Apr;21(1):81-9
- Wu Y, Marsh JW “Selective transcription and modulation of resting T cell activity by preintegrated HIV DNA” *Science* 2001 Aug 24;293(5534):1503-6
- Fanales-Belasio E, Moretti S, Nappi F, Barillari G, Micheletti F, Cafaro A, Ensoli B: “Native HIV-1 Tat protein targets monocyte-derived dendritic cells and enhances their maturation, function and antigen-specific T cell responses” *J Immunol*. 2002 Jan 1;168(1):197-206
- Corinti S, Chiarantini L, Dominici S, Laguardia ME, Magnani M, Girolomoni G: “Erythrocytes deliver Tat to interferon-gamma-treated human dendritic cells for efficient initiation of specific type 1 immune responses in vitro” *J Leukoc Biol*. 2002 Apr;71(4):652-8
- Dominici S, Laguardia ME, Serafini G, Chiarantini L, Fortini C, Tripiciano A, Brocca-Cofano E, Scoglio A, Caputo A, Fiorelli V, Gavioli R, Cafaro A, Ensoli B, Magnani M: “Red blood cell-mediated delivery of recombinant HIV-1 Tat protein in mice induces anti-Tat neutralizing antibodies and CTL” *Vaccine*. 2003 May 16;21(17-18):2073-81