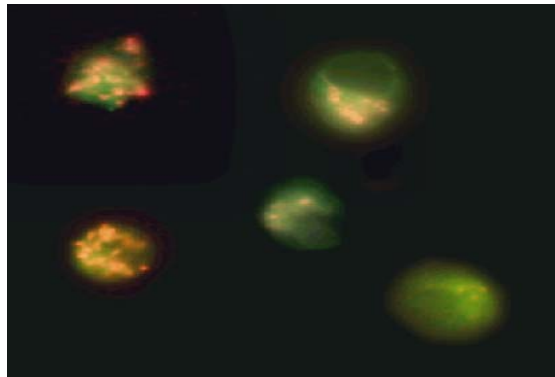


MitoPT™ JC-1

Mitochondrial Permeability Transition Detection Kits

MitoPT™ JC-1 100 Test Kit – catalog # 924

MitoPT™ JC-1 400 Test Kit – catalog # 911



Cells bearing healthy/polarized mitochondria appear red
Cells bearing depolarized mitochondria appear green



For technical questions and orders, please contact us at:

1-800-829-3194
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952-888-8988 fax
www.immunochemistry.com

Ordering Information

MitoPT™ JC-1 Kit – 100 tests/kit – catalog # 924

MitoPT™ JC-1 Kit – 400 tests/kit – catalog # 911

Related Products

MitoPT™ **TMRE $\Delta\Psi_m$ 100** Kit – 100 tests/kit – catalog # 9102

MitoPT™ **TMRE $\Delta\Psi_m$ 500** Kit – 500 tests/kit – catalog # 9103

MitoPT™ **TMRM $\Delta\Psi_m$ 100** Kit – 100 tests/kit – catalog # 9104

MitoPT™ **TMRM $\Delta\Psi_m$ 500** Kit – 500 tests/kit – catalog # 9105

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Table of Contents

1. Introduction	4
2. Contents of the MitoPT™ JC-1 Kit	5
3. Recommended Materials and Equipment	6
4. Instrumentation	6
5. Storage and Shelf-Life	6
6. Safety Information	6
7. Overview of the MitoPT™ JC-1 Protocol	6
8. Induction of Apoptosis	7
9. Preparation of 1X Assay Buffer	7
10. Reconstitution of the 100X MitoPT™ JC-1 Stock	7
11. Preparation of 1X MitoPT™ JC-1 Solution for Immediate Use	8
12. Storage of 100X MitoPT™ JC-1 Stock for Future Use	9
13. Preparation of 1X MitoPT™ JC-1 Solution from a Frozen Aliquot ...	9
14. Flow Cytometry Staining Protocol	10
15. Flow Cytometer Set Up	11
16. Single-Parameter Analysis Using a Flow Cytometer	11
17. Multi-Parameter Analysis Using a Flow Cytometer	11
18. Flow Cytometry Sample Data	11
19. 96-Well Fluorescence Spectroscopy Staining Protocol	13
20. 96-Well Fluorescence Plate Reader Set Up	14
21. 96-Well Fluorescence Spectroscopy Sample Data	15
22. Fluorescence Microscopy Staining Protocol for Adherent Cells.....	16
23. Fluorescence Microscopy Staining Protocol for Suspension Cells .	17
24. Fluorescence Microscope Set Up	18
25. Fluorescence Microscopy Sample Data	18
26. References	19

1. Introduction

Detection of the mitochondrial permeability transition event (PT) provides an early indication of the initiation of cellular apoptosis. This process is typically defined as a collapse in the electrochemical gradient across the mitochondrial membrane, as measured by the change in the membrane potential ($\Delta\Psi_m$).

Changes in the mitochondrial $\Delta\Psi_m$ lead to the insertion of proapoptotic proteins into the membrane and possible oligomerization of BID, BAK, BAX or BAD. This could create pores, which dissipate the transmembrane potential, thereby releasing cytochrome c into the cytoplasm (1-5).

Loss of mitochondrial $\Delta\Psi_m$, indicative of apoptosis, can be detected by a unique fluorescent cationic dye, 5,5',6,6'-tetrachloro-1,1',3,3'-tetraethylbenzamidoazolocarbocyanin iodide, commonly known as JC-1 (6). This dye has been incorporated into the user-friendly MitoPT[™] JC-1 kit for the simple and reproducible detection of the PT event in apoptotic cells.

The structure of ImmunoChemistry Technologies' MitoPT[™] JC-1 reagent allows it to easily penetrate cells and healthy mitochondria. Once inside a healthy non-apoptotic cell, the lipophilic MitoPT[™] JC-1 reagent, bearing a delocalized positive charge, enters the negatively charged mitochondria where it aggregates and fluoresces red (7). These aggregates, first described by Jelley in 1937, are referred to as J-aggregates (8). When the mitochondrial $\Delta\Psi_m$ collapses in apoptotic cells, the MitoPT[™] JC-1 reagent can no longer accumulate inside the mitochondria. Instead, it is dispersed throughout the cell in a monomeric form, which fluoresces green (9). Use of the MitoPT[™] JC-1 kit allows the easy distinction between healthy/ non-apoptotic red fluorescent cells and apoptotic or mitochondrial membrane compromised green fluorescent cells.

The MitoPT[™] JC-1 kit can be used in conjunction with your existing apoptosis or metabolic stress protocols. Grow your cells in culture and induce apoptosis according to your existing procedure (also reserve a non-induced population of cells as a negative control). Once you have induced apoptosis or mitochondrial depolarization activity in your cells, add the 1X MitoPT[™] JC-1 solution to each population and incubate the cells for an additional 15 minutes. During this incubation time, the JC-1 reagent will enter each cell and the mitochondria inside. If the cell is not undergoing apoptosis or oxidative/metabolic stress, the mitochondrial $\Delta\Psi_m$ will remain intact, and the JC-1 reagent will concentrate and form J-aggregates inside the mitochondria and fluoresce red. If the cell is apoptotic, the mitochondrial $\Delta\Psi_m$ will be breaking down, thereby causing the JC-1 reagent to be dispersed throughout the entire cell and fluoresce green.

The MitoPT[™] JC-1 reagent excites at 488-490 nm. The monomeric dye structure emits at 527 nm, whereas the J-aggregates in healthy (non-apoptotic) mitochondria emit at 590 nm (7).

The MitoPT[™] JC-1 kit can detect the mitochondrial depolarization event using three different technologies: flow cytometers; fluorometric plate readers; and fluorescence microscopes.

Following the flow cytometer and fluorescence microscope protocols, each sample to be stained requires only 0.5 mL of 1X MitoPT[™] JC-1 solution (equal to 5 μ L of 100X MitoPT[™] stock). The MitoPT[™] JC-1 100 Kit will test 100 samples; the MitoPT[™] JC-1 500 Kit will test 500 samples. Following the fluorescence plate reader protocol, each sample requires 1 mL of 1X MitoPT[™] JC-1 solution (requiring 10 μ L of 100X MitoPT[™] JC-1 stock). The MitoPT[™] JC-1 100 Kit will test 50 samples; the MitoPT[™] JC-1 400 Kit will test 200 samples.

When cells stained with MitoPT[™] JC-1 are evaluated on a flow cytometer, the instrument will measure loss of $\Delta\Psi_m$ by monitoring the reduction of red fluorescence in cell populations where the proton gradient across the inner mitochondrial membrane is diminished. Healthy cells which contain mitochondria that are maintaining a normal proton and pH gradient across the inner mitochondrial membrane, will continue to concentrate the JC-1 dye and exhibit the classic orange-red fluorescence. Experimental induction of apoptosis or metabolic stress conditions will trigger an increase in the overall level of mitochondrial depolarization. This leads to a dramatic reduction in JC-1 J-aggregate formation in the effected mitochondria. Monomeric JC-1 which is not in the J-aggregate form exhibits a green fluorescence (see Section 18 for sample data).

When cells stained with MitoPT[™] JC-1 are analyzed with a fluorescence plate reader, the instrument will measure apoptosis by monitoring the amount of red fluorescence. Healthy cells will give a high relative fluorescence unit (RFU) emissions output of red fluorescence. As the mitochondrial $\Delta\Psi_m$ collapses, indicating an apoptosis induction event, an increasing number of cells will give a lower red fluorescence RFU reading as the dispersed JC-1 dye converts to a green fluorescing monomeric form (see Section 21 for sample data).

Looking at the cells under a fluorescence microscope, non-apoptotic cells will appear to have orange-red fluorescent mitochondrial structures (the JC-1 dye aggregates) within healthy mitochondria. In contrast, apoptotic cells will appear mostly green. Cells still undergoing apoptosis will contain less and less red fluorescing mitochondria. Eventually, the entire cell will appear more and more green as the mitochondrial $\Delta\Psi_m$ dissipates (see Section 25 for sample data).

2. Contents of the MitoPT[™] JC-1 Kit:

- MitoPT[™] JC-1 Reagent, lyophilized, part# 6261 (100) or 6260 (400).
- 10X Assay Buffer, part# 6161 (30 mL), or 6259 (125 mL).
- 50 mM Carbonylcyanide *m* - chlorophenylhydrazone (CCCP) concentrate in DMSO, 1 x (125 μL vial), part# 6257 in 100 test size kit or 1 x (600 μL vial), part# 6258 in 400 test size kit.
- Assay Manual with protocols for 3 applications: Flow Cytometer; Fluorometer; and Fluorescence Microscope.

3. Recommended Materials and Equipment (not all are required):

- Cultured cells and media
- Protocol and reagents to induce metabolic stress, mitochondrial depolarization, or apoptosis
- 15 mL polystyrene centrifuge tube (1 per sample)
- Microfuge at 13,000 X g, Clinical centrifuge at 100 – 1,000 X g
- Pipette(s) capable of dispensing at 10 μL, 500 μL, and 1 mL
- Graduated cylinder
- Dimethyl Sulfoxide - DMSO (125 μL or 500 μL)
- Vortexer
- Amber vials or polypropylene tubes for storage at –20°C
- 37°C CO₂ incubator
- Hemocytometer

4. Instrumentation (not all are required):

- Flow cytometer with excitation between 488 - 490nm, and emission at 527 and 590 nm.
- 96-well fluorescence plate reader with excitation between 488 - 490 nm, and emission at 590 nm (dual emission at 527 nm and 590 - 600 nm is best), with endpoint reading and black round or flat bottom 96-well microtiter plates.
- Fluorescence microscope with broad band path filters and slides.

5. Storage and Shelf-Life

- Store the unopened kit (and each unopened component) at 2°C to 8°C until the expiration date.
- Once reconstituted, store the 100X or 200X MitoPT[™] JC-1 stock at –20°C for 6 months. It may be frozen twice.
- Once diluted, store the 1X assay buffer at 2°C to 8°C up to 7 days.

6. Safety Information

- Use gloves while handling the MitoPT[™] JC-1 reagent, and the 10X assay buffer.
- Dispose of all liquid components down the sink and flush with copious amounts of water. Solid components may be tossed in standard trash bins.
- MSDS are available at www.immunochemistry.com or by calling 1-800-829-3194.

7. Overview of the MitoPT[™] JC-1[™] Protocol

Staining cells with MitoPT[™] JC-1[™] takes only about 15 minutes.

However, the MitoPT[™] JC-1[™] kit is used with living cells, which may take several hours to prepare. In addition, once the cells are grown, they must be induced to undergo apoptosis or oxidative stress, that may also take several hours to complete. As the 1X MitoPT[™] JC-1[™] solution must be used immediately, prepare the MitoPT[™] reagents at the end of your metabolic stress or apoptosis induction process. Here is a quick overview of the MitoPT[™] JC-1[™] protocol:

1. Culture cells to a density optimal for apoptosis induction according to your specific induction protocol, but not to exceed **10⁶ cells/mL**.
2. At the same time, culture a non-induced negative control cell population (at the same density as the induced population).
3. Induce apoptosis or mitochondrial depolarization following your protocol (2 sample protocols are mentioned in Section 8).
4. Prepare 1X assay buffer and warm to 37°C (Section 9).
5. Prepare 100X or 200X MitoPT[™] JC-1[™] stock (Section 10).
6. Prepare 1X MitoPT[™] JC-1[™] solution (Section 11 or 13).
7. Stain cells with 1X MitoPT[™] JC-1[™] solution (Section 14, 19, 22, or 23).
8. Analyze data (Section 15, 20, or 24).

8. Induction of Apoptosis or Mitochondrial Depolarization

The MitoPT[™] JC-1[™] kit work is compatible with any apoptosis or mitochondrial assessment protocol. An apoptotic positive control can routinely be established in Jurkat leukemic T cells using either a 2 µg/ml camptothecin or 1 µM staurosporine induction for 3-4 hours at 37°C. Depolarized mitochondria containing (positive control) cell populations can easily be prepared by incubating cells with 5-50 µM CCCP for 30 to 60 minutes at 37°C. (50 mM CCCP stock included in kit). This proton gradient uncoupling agent quickly reduces the electrochemical potential across the inner mitochondrial membrane, resulting in a rapid intracellular mitochondrial depolarization event (10-11). To generate a CCCP-induced positive control population, follow the brief protocol listed below:

1. Pre-determine when you intend to run the mitochondrial depolarization analysis on your control and experimental (treatment) cell populations. Approximately 75 minutes prior to performing this analysis using the MitoPT[™] JC-1[™] kit, plan to generate a positive and negative mitochondrial depolarization control sample set using the CCCP mitochondria depolarizing agent that is included in this kit.
2. Gently warm the 50 mM CCCP stock reagent vial to RT. Be sure to mix or gently vortex the CCCP stock vial contents prior to adding it to cell cultures.
3. Spike the designated and labeled positive control population with enough CCCP de-polarizing reagent to obtain a 10–50 µM CCCP concentration in the cell culture media. If using a 50 µM CCCP

concentration for mitochondrial depolarization, simply spike in 1 μ L of 50 mM CCCP stock per mL of cell suspension/overlay media. If using a lower concentration for induction, make a 1:10 dilution of an aliquot of the 50 mM CCCP stock in tissue culture grade DMSO, so as to be able to accurately dispense the CCCP into the cell culture tube or flask.

4. Prepare a negative control cell population that was spiked with the same volume of tissue culture grade DMSO as was used to spike the CCCP cell population.
5. Incubate the negative and positive control cell populations for 30–60 minutes at 37°C in the CO₂ incubator to allow for the depolarization process to proceed.

9. Preparation of 1X Assay Buffer

The assay buffer is formulated for use as reaction buffer, and for washing the cells. It is supplied as a 10X concentrate which must be diluted to 1X with DI H₂O prior to use.

If cells can not be analyzed within a 2-3 hour time frame, it is recommended that a reagent grade BSA powder be added to the 1X Assay Buffer to a 1% w/v final concentration. This will assure an extended cell viability environment until analysis can be completed.

1. If necessary, gently warm the 10X concentrate to completely dissolve any salt crystals that may have come out of solution.
2. Dilute the 10X assay buffer concentrate 1:10 in DI H₂O. For example, add the contents of one 30 mL bottle to 270 mL of DI H₂O. This will yield a 300 mL volume of 1X Assay Buffer for use as a wash or readout suspension medium. For smaller volumes, add 10 mL 10X assay buffer to 90 mL DI H₂O to yield 100 mL of 1X Assay Buffer.
3. Stir the 1X solution for at least 5 minutes.
4. If not using the 1X assay buffer the same day it was prepared, store it at 2°C to 8°C for up to 7 days. Warm the 1X Assay Buffer to RT prior to use.

10. Reconstitution of the 100X or 200X MitoPT[™] JC-1 Stock

The MitoPT[™] JC-1 dye reagent is supplied as a highly concentrated lyophilized powder. It must first be reconstituted, forming a 100X (100 test size vial) or 200X (400 test size vial) stock concentrate, and then diluted 1:100 or 1:200 respectively, to form a final 1X working solution. The 1X working solution must be prepared immediately prior to use; however, the reconstituted 100X or 200X stock can be stored at –20°C for 6 months, and used twice during that time.

- **The newly reconstituted 100X or 200X MitoPT[™] JC-1 stock must be used or frozen immediately after it is prepared and protected from light during handling.**

1. For the 100 test kit, reconstitute the 100-test vial (part# 6261) with 500 μ L DMSO at room temperature (RT), forming a **100X** stock.
2. Or, for the 400 test kit, reconstitute the 400-test vial (part# 6260) with 1 mL DMSO at room temperature (RT), forming a 200X stock.
3. Re-cap the vial and invert it several times to fully dissolve the MitoPT[™] JC-1 dye reagent.
4. Immediately use the 100X stock by diluting it to 1X (Section 11).
5. Or, aliquot and store it at -20°C (Section 12).

11. Preparation of 1X MitoPT[™] JC-1 Solution for Immediate Use

Using the freshly reconstituted 100X or 200X MitoPT[™] JC-1 stock, prepare the 1X working strength MitoPT[™] JC-1 solution by diluting the stock 1:100 or 1:200 respectively with 37°C 1X assay buffer (or, you may substitute your own cell culture media, warmed to 37°C , in place of the 1X assay buffer). Each sample to be stained requires only 0.5 mL of 1X MitoPT[™] JC-1 solution (equal to 5 μ L of 100X or 2.5 μ L of 200X MitoPT[™] JC-1 stock).

● The 1X working strength MitoPT[™] JC-1 solution must be used immediately – prepare it after apoptosis induction is completed.

1. Warm the 1X assay buffer (or cell culture media) to 37°C .
2. For the 100-test kit, add 500 μ L of the 100X MitoPT[™] JC-1 stock from each vial to 49.5 mL of the 37°C 1X assay buffer or cell culture media.
3. Or, for the 400-test kit, add 1 mL of the 200X MitoPT[™] JC-1 stock to 199 mL of the 37°C 1X assay buffer or cell culture media.
4. Or, if not using the entire vial of 100X or 200X MitoPT[™] stock, you could for example, dilute 10 μ L of 100X MitoPT[™] stock to 990 μ L of 1X assay buffer or cell culture media.
5. Vortex the 1X working strength MitoPT[™] JC-1 solution thoroughly.
6. If particulate matter is present, the solution should be clarified by centrifugation at 13,000 X g in a microfuge for 3 minutes, or 20 minutes in a clinical centrifuge at $>1,000$ X g at RT. **This is especially important when using a fluorescence plate reader or taking fluorescence microscopy photos for publication.**

● Clarification of the 1X solution is absolutely necessary when performing the fluorescence plate reader protocol. Any free dye crystals in the solution will interfere with the OD reading, leading to a falsely increased reading of red fluorescence. The fluorescent JC-1 precipitate is easily gated out during the initial flow cytometry setup and will not interfere with flow cytometer analysis.

7. If centrifuged, transfer the clarified supernatant to a clean tube and discard particulates.

8. Go on to the staining protocol (Section 14, 19, 22, or 23).

● **To avoid photo-bleaching and degradation of the dye, protect the MitoPT[™] JC-1 dye reagent from light while handling.**

12. Storage of 100X or 200X MitoPT[™] JC-1 Stock for Future Use

If not all of the 100X or 200X MitoPT[™] JC-1 stock will be used at the same time it is reconstituted, the unused portion may be stored at -20°C for 6 months. During that time, the 100X or 200X MitoPT[™] JC-1 stocks may be thawed and used twice. After the second thaw, discard any remaining MitoPT[™] JC-1 concentrate. If you anticipate using it more than twice, make small aliquots in amber vials or polypropylene tubes. When ready to use, follow Section 13.

13. Preparation of 1X MitoPT[™] JC-1 Solution from a Frozen Aliquot

If some of the 100X or 200X MitoPT[™] JC-1 stock was previously reconstituted and then stored at -20°C, it may be used 2 more times within 6 months. An example of the use of a **100X** stock concentrate is given below

1. Thaw the 100X MitoPT[™] JC-1 stock and protect from light.
2. Once the aliquot has become liquid, dilute the 100X MitoPT[™] JC-1 stock 1:100 in 37°C 1X assay buffer or cell culture media. For example, mix 10 µL of 100X MitoPT[™] JC-1 stock with 990 µL of 37°C 1X assay buffer or cell culture media.
3. If the 100X MitoPT[™] JC-1 stock was frozen immediately after reconstitution and never been thawed, return it to the freezer. If the stock has been thawed once before, discard it.
4. Vortex the 1X working strength MitoPT[™] JC-1 solution thoroughly.
5. If particulate matter is present, the solution must be clarified by centrifugation at 13,000 X g in a microfuge for 3 minutes, or 15 minutes in a clinical centrifuge at 1,000 X g at RT.

● **As stated above in Section 11, clarification of the 1X solution is especially important when performing fluorescence plate reader or fluorescence microscope analysis protocols that would be submitted for publication.**

6. If centrifuged, transfer the clarified supernatant to a clean tube and discard particulates.
7. Go on to the staining protocol (Section 14, 19, 22, or 23).

14. Flow Cytometry Staining Protocol

Following the flow cytometer protocols, each sample to be stained requires only 0.5 mL of 1X MitoPT[™] JC-1 solution (equal to 5 µL of 100X MitoPT[™] JC-1 stock). The MitoPT[™]-100 Kit will test 100 samples; the MitoPT[™]-400 Kit will test 400 samples.

1. As discussed in Section 7, culture cells to a density optimal for induction of apoptosis or $\Delta\Psi_m$ disruption model experiments, according to your specific cell culture protocols.
2. Induce apoptosis following your protocol.
3. At the same time, culture a non-induced negative control cell population.
4. After the induction process, transfer 0.5 mL of each cell suspension into a sterile 15 mL polystyrene centrifuge tube.

● Cell density in the cell culture flasks should not exceed 10⁶ cells per mL. Cells cultivated in excess of this concentration may begin to naturally enter apoptosis. Optimal cell concentration will vary depending on the cell line used. Density can be determined by counting cell populations on a hemocytometer.

5. Centrifuge cells at < 200 X g for 5 minutes at RT.
6. Carefully remove and discard the supernatant.
7. Gently vortex or pipette the cell pellets, to disrupt any cell-to-cell clumping.
8. Resuspend cells in 0.5 mL of RT 1X MitoPT[™] JC-1[™] solution.
9. Incubate the cells, staining with the MitoPT[™] JC-1[™] dye reagent, at 37°C for 10-15 minutes in a CO₂ incubator.
10. Warm the 1X assay buffer to 37°C (Section 9).
11. Add 2 mL of 1X assay buffer to each tube.
12. Mix each tube.
13. Centrifuge the stained cells at < 200 X g for 5 minutes at RT.
14. Carefully remove and discard supernatant.
15. Gently vortex the pellet to disrupt any cell-to-cell clumping.
16. Resuspend the cells in 1 mL of 1X assay buffer.
17. Centrifuge the stained cells at < 200 X g for 5 minutes at RT.
18. Carefully remove and discard supernatant.
19. Gently vortex the pellet to disrupt any cell-to-cell clumping.
20. Resuspend the cells in 1 mL of 1X assay buffer.
21. Centrifuge the stained cells at < 200 X g for 5 minutes at RT.
22. Carefully remove and discard supernatant.
23. Gently vortex the pellet to disrupt any cell-to-cell clumping.
24. Resuspend the cell pellet in 0.5 mL of 1X assay buffer.
25. Analyze cells by flow cytometry (Sections 15-18).

15. Flow Cytometer Set Up

Healthy cells containing the aggregated MitoPT[™] JC-1[™] dye reagent within their mitochondria (which fluoresces red) can be detected in the FL2 channel. Apoptotic cells, which contain non-aggregated green MitoPT[™] JC-1[™] monomers, can be detected in the FL1 channel.

16. Single-Parameter Analysis Using a Flow Cytometer

1. Generate a log FL2 (X-axis) versus cell count.

2. Adjust the FL2 PMT voltage to allow the peak to fall within the third log decade when running the negative control sample.
3. When running the induced positive samples, use the same adjusted PMT voltage as was determined for the negative control.
4. Observe the mean fluorescence of both the apoptotic positive cell population, as well as the mean fluorescence of the treated negative control cell population.

● **The apoptotic cell population displays a lower red fluorescence signal intensity (FL2 axis) than the negative control population.**

17. Multi-Parameter Analysis Using a Flow Cytometer

1. Create a log FL1 (X-axis) versus log FL2 (Y-axis) scatter plot.
2. Add two regions, R2 and R3, as shown in Figure 1.
3. Run the negative control sample first (non-induced cells). Adjust the FL1 and FL2 PMT voltages so that the majority of the cell population falls within the upper right hand region. The peak of the dual fluorescent population should fall within the second and third log decade scale of FL1 (X-axis) and FL2 (Y-axis) as seen in Figure 1.
4. Adjust R2 so that greater than 95% of the dual fluorescent cell population falls within this region. The number of cells falling into this region will vary depending on the condition of the culture and cell type.
5. Next, adjust R3 so that it falls directly below R2, as in Figure 1.
6. Using the same PMT settings established for the non-induced negative control sample in Step 3, run the induced positive sample. If a change in mitochondrial $\Delta\Psi_m$ has occurred, an increase in the number of cells falling in R3 is observed as shown in Figure 2.

● **This reflects a reduction in red fluorescence.**

7. If the induced sample exhibits only a minimal change in red emission, increase the FL2-FL1 compensation and repeat Steps 3-6.

18. Flow Cytometry Sample Data

When cells stained with MitoPT[™] JC-1 are run through a flow cytometer, the instrument will measure apoptosis by monitoring the amount of red fluorescence in each region. Healthy cells, which fluoresce red, will appear in R2. As the mitochondrial $\Delta\Psi_m$ collapses and cells enter apoptosis or some other oxidative stress mechanism, the amount of red fluorescence will drop. An increasing number of cells will fall into R3 corresponding to a loss of red fluorescence as the dispersed MitoPT[™] JC-1 dye converts to a monomeric form and fluoresces green.

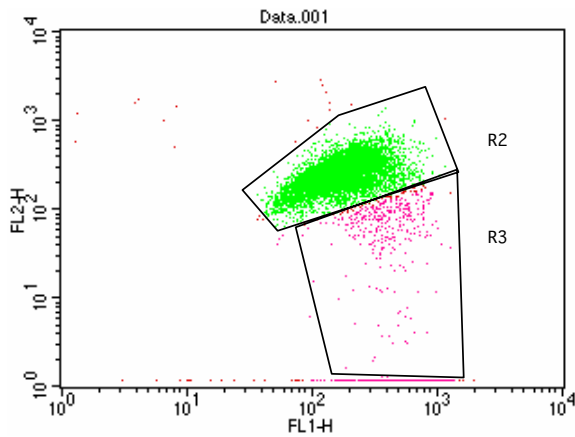


Figure 1: Negative cell control, Log FL1 (X-axis) and log FL2 (Y-axis) R2 = 91.07% R3 = 8.08%.

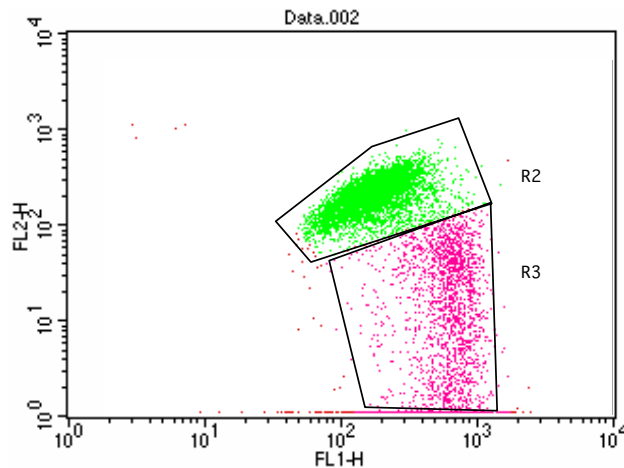


Figure 2: Positive cell control, Log FL1 (X-axis) and log FL2 (Y-axis) R2 = 60.93% R3 = 38.13%.

In Figures 1 and 2, cells were analyzed in a FACS Caliber Becton Dickinson Flow Cytometer. Jurkat cells were either treated with DMSO (negative, non-induced cells) or with staurosporine (apoptotic, induced cells) for 3 hours at 37°C and then labeled with MitoPT[™] JC-1 for 15 minutes. Collapse of the mitochondrial $\Delta\Psi_m$ is indicated by an increase in the number of cells falling into R3 corresponding to a loss of red fluorescence, indicative of the onset of apoptosis or other $\Delta\Psi_m$ depolarizing event. (In these figures of flow cytometer data, non-apoptotic cells are in R2, which prints in green, and apoptotic cells are in R3, which prints in pink.)

19. 96-Well Fluorescence Spectroscopy Staining Protocol

Following the fluorescence plate reader protocol, each sample requires 1 mL of 1X MitoPT[™] JC-1 solution (equal to 10 μ L of 100X MitoPT[™] JC-1 stock). The MitoPT[™]-JC-1 100 Kit will test 50 samples; the MitoPT[™]-JC-1 400 Kit will test 200 samples.

1. As discussed in Section 7, culture cells to a density optimal for your apoptosis or oxidative/metabolic stress induction experiments, according to your specific cell culture protocol.
- **Cell density in the cell culture flasks should not exceed 10⁶ cells per mL. Cells cultivated in excess of this concentration may begin to naturally enter apoptosis. Optimal cell concentration will vary depending on the cell line used. Density can be determined by counting cell populations on a hemocytometer.**
2. Generate your experimental apoptosis or $\Delta\Psi_m$ disrupted cells using your preferred protocol
 3. At the same time, culture an equal volume of non-induced cells for a negative control cell population. Make sure that both tubes of cells contain similar quantities of cells.
 4. Pellet cells by centrifugation at < 200 X g for 5 minutes at RT.
- **When concentrated, cells should have been grown to yield a 0.5 mL concentrated pool between 1 - 2 X 10⁶ cells/mL.**
5. Carefully remove and discard the supernatants.
 6. Resuspend at between 0.5 – 2 X 10⁶ cells in 1 mL of 1X working strength MitoPT[™] JC-1 solution (Sections 11 and 13).
 7. Gently vortex or pipette the cell pellets to disrupt any cell-to-cell clumping.
 8. Incubate the cells (staining with the MitoPT[™] JC-1 dye reagent) at 37°C for 10-15 minutes in a CO₂ incubator.
 9. Warm the working strength 1X assay buffer to 37°C (Section 9).
 10. Add 2 mL of 1X assay buffer to each tube.
 11. Mix each tube.
 12. Centrifuge the stained cells at < 200 X g for 5 minutes at RT.
 13. Carefully remove and discard supernatants.
 14. Gently vortex the pellets to disrupt any cell-to-cell clumping.
 15. Resuspend the cells in 1 mL of 1X assay buffer.
 16. Take out a small aliquot to determine the concentration of both the induced and non-induced cell populations.
 - a. Remove a 50 μ L aliquot of each cell population.
 - b. Add to 450 μ L PBS (forming a 1:10 dilution of each).
 - c. Count the cells (a hemocytometer may be used).

- d. After counting, compare the density of each. The non-induced population may have more cells than the induced population, as some induced cells may be lost during the apoptotic process. If necessary, adjust the volume of the induced cell suspension to match that of the non-induced suspension (Step 20).
17. Centrifuge the remaining stained cells at 400 X g for 5 minutes at RT.
 18. Carefully remove and discard supernatants.
 19. Gently vortex the pellets to disrupt any cell-to-cell clumping.
 20. Adjust the volume of the induced cell suspension to match that of the non-induced suspension.

● **A minimum of 1×10^5 cells/well is recommended to generate an adequate fluorescence signal using most 96-well plate readers.**

- a. Resuspend the non-induced cell pellets in 500 μ L - 1 mL of 1X assay buffer to produce a cell suspension at least 1×10^6 cells/mL. The volume may vary depending upon cell density. You may find that you can get by using a lower cell number per well in your particular cell model system and fluorescence detection plate reader but this should be pre-determined for each individual case.
 - b. Resuspend the induced cell population in a volume of 1X assay buffer to yield the same concentration of cells as the non-induced cell suspension.
21. For each sample to be tested, dispense 100 μ L into each of 2 - 4 wells in a **black** round or flat bottom 96-well microtiter plate.

● **The 96-well microtiter plate used to analyze the cells must be black. A conventional clear plate will severely diminish the assay sensitivity.**

22. Analyze cells using a fluorescence plate reader (Section 20).

20. 96-Well Fluorescence Plate Reader Set Up

1. Set the plate reader to perform an endpoint read.
2. Set the excitation wavelength at 488 - 490 nm
3. Set the emission wavelengths to 527 nm for green fluorescence and 590 - 600 nm for red fluorescence. If your plate reader cannot read dual emission wavelengths at the same time, use the red fluorescence setting of 590 - 600 nm to perform the analysis.
4. Read the samples and print out a copy of the fluorescence (RFU) output. Save the data electronically for future reference.

21. 96-Well Fluorescence Spectroscopy Sample Data

When cells stained with MitoPT[™] JC-1 are analyzed with a fluorescence plate reader, the instrument will measure the amount of orange-red fluorescence. Healthy cells will give a high OD reading of red fluorescence; apoptotic or metabolically stressed cells will exhibit a reduced level of orange-red fluorescence intensity.

By comparing the average 590-600 nm OD signal in stimulated versus non-stimulated sample wells, loss of $\Delta\Psi_m$ can be monitored. As the $\Delta\Psi_m$ collapses (commonly an early stage of apoptosis) and the JC-1 dye aggregates transition to the green fluorescing monomeric form, more and more cells will lose their orange-red fluorescence (Figures 3 and 4).

Using the dual fluorescence characteristic of the dye, the changes in the mitochondrial $\Delta\Psi_m$ can be most accurately assessed by comparing the ratios of 590-600 nm (red) / 527 nm (green) ODs. When apoptosis is induced, the red/green OD ratio drops compared to the negative (non-stimulated) control wells. In other words, the red OD signal decreases and the green OD signal remains constant or increases. This drop in orange-red signal corresponds to a reduction in the number of healthy mitochondria able to maintain the negative potential necessary to concentrate the JC-1 dye in the red fluorescing, aggregate form.

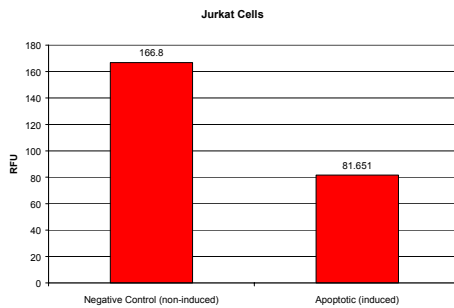


Figure 3: Red fluorescence of Jurkat cells.

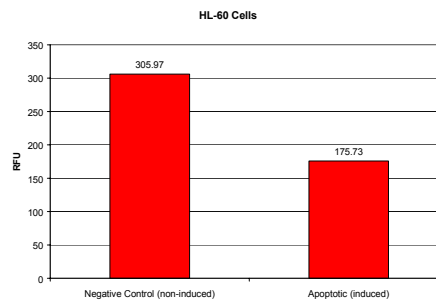


Figure 4: Red fluorescence of HL-60 cells.

In Figures 3 and 4, the cells were either treated with DMSO (negative, non-induced cells – bars on the left side of each graph) or with staurosporine (apoptotic, induced cells - bars on the right side of each graph) for 4 hours and then labeled with the 1X MitoPT[™] JC-1 solution for 15 minutes. The samples were then read on a 96-well fluorescence plate reader using the settings described above. As the mitochondrial $\Delta\Psi_m$ collapses, indicating apoptosis, the amount of red fluorescence drops by 51% in the Jurkat cells and 43% in HL-60 cells.

If there are free aggregates of the MitoPT[™] JC-1 dye in solution with the cells, they will fluoresce red and interfere with the data, leading to a falsely increased reading of red fluorescence (Sections 11 and 13). **This is why it is essential to pre-centrifuge the 1X JC-1 dye solution prior to staining cells that will be evaluated on a fluorescence plate reader.**

22. Fluorescence Microscopy Staining Protocol for Adherent Cells

Following the fluorescence microscope protocol, each sample to be stained requires only 0.5 mL of 1X MitoPT[™] JC-1 solution (equal to 5 μ L of 100X MitoPT[™] JC-1 stock). The MitoPT[™] JC-1 100 Kit will test 100 samples; the MitoPT[™] JC-1 400 Kit will test 400 samples.

1. Culture cells on a sterile coverslip or chamberslide to a cell density optimal for apoptosis induction according to your specific induction protocol.

● Cell density should not exceed the threshold where cell sloughing occurs.

2. As discussed in Section 8, induce apoptosis following your protocol.
3. At the same time, culture an equal volume of non-induced cells for a negative control cell population.
4. Make sure that both the negative control and induced positive cell populations contain similar quantities of cells.
5. Remove the media from induced and non-induced monolayer cultures.
6. Add enough 1X MitoPT[™] JC-1 solution (Sections 11 and 13) to cover the cells on the slide.
7. Incubate the cells, staining with the MitoPT[™] JC-1 dye reagent, at 37°C for 15 minutes in a CO₂ incubator.
8. Warm the 1X assay buffer to 37°C (Section 9).
9. Carefully remove and discard staining media.
10. Wash the monolayer cultures with 1 to 2 mL of 1X assay buffer.
11. Discard wash.
12. Add a drop of 1X assay buffer plus coverslip.
13. Examine using a fluorescence microscope (Sections 24 and 25).

23. Fluorescence Microscopy Staining Protocol for Suspension Cells

Following the fluorescence microscope protocol, each sample to be stained requires only 0.5 mL of 1X MitoPT[™] JC-1 solution (equal to 5 μ L of 100X MitoPT[™] JC-1 stock). The MitoPT[™] JC-1 100 Kit will test 100 samples; the MitoPT[™] JC-1 400 Kit will test 400 samples.

1. Culture cells to a cell density optimal for apoptosis induction according to your specific induction protocol.

- **Optimal cell densities will vary with the cell line; cell concentrations may be determined using a hemocytometer.**
2. As discussed in Section 8, induce apoptosis following your protocol.
 3. At the same time, culture an equal volume of non-induced cells for a negative control cell population.
 4. Make sure that both the negative control and induced positive cell populations contain similar quantities of cells.
 5. Once induced, count cells (a hemocytometer may be used).
 6. Transfer 1-2 X 10⁶ cells to a centrifuge tube.
 7. Centrifuge cells at < 200 X g for 5 minutes at RT.
 8. Carefully remove and discard the supernatants.
 9. Gently vortex or pipette the cell pellets, to disrupt any cell-to-cell clumping.
 10. Warm the 1X assay buffer (or cell culture media) to 37°C.
 11. For the 100-test kit, add 500 µL of the 100X MitoPT[™] JC-1 stock from each vial to 49.5 mL of the 37°C 1X assay buffer or cell culture media.
 12. Or, for the 400-test kit, add 1 mL of the 200X MitoPT[™] JC-1 stock to 199 mL of the 37°C 1X assay buffer or cell culture media.
 13. **Or, if not using the entire vial of 100X or 200X MitoPT[™] JC-1 stock, you could for example, dilute 10 µL of 100X MitoPT[™] JC-1 stock to 990 µL of 1X assay buffer or cell culture media.**
 14. Vortex the 1X working strength MitoPT[™] JC-1 solution thoroughly.
 15. If particulate matter is present, the solution should be clarified by centrifugation at 13,000 X g in a microfuge for 3 minutes, or 20 minutes in a clinical centrifuge at >1,000 X g at RT. **This extra clarification step is especially important when taking publication quality fluorescence microscopy photos.**
 16. Resuspend cells in 0.5 mL 1X MitoPT[™] JC-1 solution (Sections 11 and 13).
 17. Incubate the cells (which are now being stained with the MitoPT[™] JC-1 dye reagent) at 37°C for 10 to 15 minutes in a CO₂ incubator.
 18. Warm the 1X assay buffer to 37°C (Section 9).
 19. Resuspend cells in 2 mL 1X assay buffer.
 20. Centrifuge cells at < 200 X g for 5 minutes at RT.
 21. Carefully remove and discard the supernatants.
 22. Gently vortex the cell pellets to disrupt any cell-to-cell clumping.
 23. Resuspend cells in 1 mL of 1X assay buffer.
 24. Centrifuge the stained cells at < 200 X g for 5 minutes at RT.
 25. Carefully remove and discard the supernatants.
 26. Gently vortex the cell pellets to disrupt any cell-to-cell clumping.
 27. Resuspend cells in 1 mL of 1X assay buffer.
 28. Centrifuge the stained cells at < 200 X g for 5 minutes at RT.
 29. Carefully remove and discard the supernatants.
 30. Resuspend cells in 0.5 mL 1X assay buffer.
 31. Examine a drop of cell suspension under a cover slip using a fluorescence microscope (Sections 24 and 25).

24. Fluorescence Microscope Set Up

This scope should contain a long band path emissions filter (Ex 490 nm, Em >510 nm) capable of detecting both fluorescein and rhodamine fluorescence simultaneously.

25. Fluorescence Microscopy Sample Data

The MitoPT[™] JC-1 dye will be concentrated in the mitochondria of healthy cells, thereby creating orange-red fluorescent mitochondria staining within the cell. The JC-1 dye will become evenly dispersed throughout apoptotic cells and cells where the electrochemical inner mitochondrial gradient has been compromised. Mitochondria in these cells will not concentrate the JC-1 dye which will result in an even distribution of the green fluorescing monomeric form of this dye throughout the lumen of the cell. Cells in varying stages of apoptosis will contain less and less orange-red fluorescing J-aggregate and appear mostly green (See Figure 5 below).

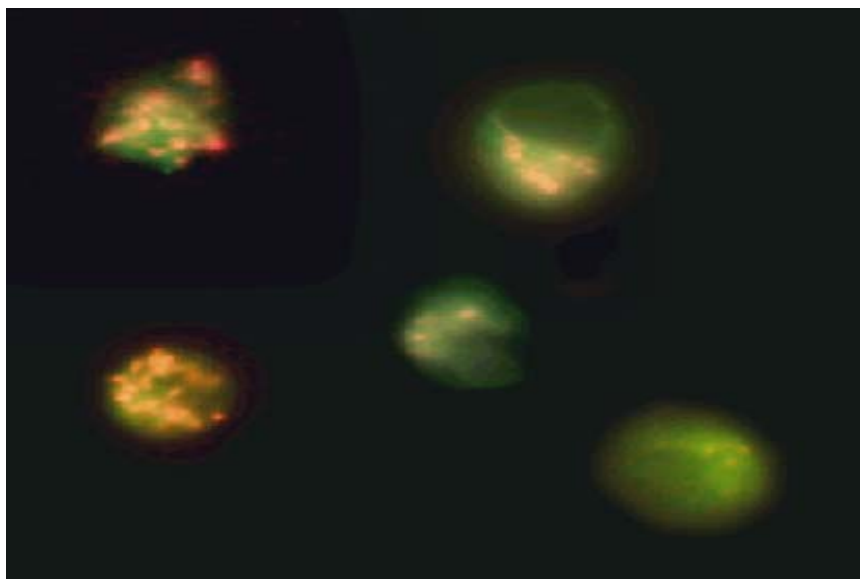


Figure 5: Jurkat cells were stained with MitoPT[™] JC-1 and viewed through a fluorescence microscope using a broad band path emissions filter. Non-apoptotic cells exhibit orange-red stained mitochondria (2 cells at left). Apoptotic cells at varying stages of mitochondrial $\Delta\Psi_m$ appear green (3 cells at right).

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