



RODENT THYROID STIMULATING HORMONE (TSH) ELISA TEST KIT

PRODUCT PROFILE AND INSTRUCTIONS

The Rodent TSH ELISA test is an immunoassay designed for the quantitative determination of thyroid stimulating hormone (TSH) in serum/plasma samples of rat/mouse and related species.

TEST PRINCIPLE:

The Rodent TSH ELISA test is based on the principle of a solid phase enzyme-linked immunosorbent assay. The assay system utilizes affinity purified antibody directed against intact rodent TSH molecule for solid phase (microtiter wells) immobilization and a mouse anti-TSH antibody is in the antibody-enzyme (horseradish peroxidase) conjugate. The test sample is allowed to react simultaneously with the two antibodies, resulting in the TSH molecules being sandwiched between the solid phase and enzyme-linked antibodies. After a 3 hours of incubation period at 37 °C, the wells are washed with wash solution to remove unbound-labeled antibodies. A solution of TMB is added and incubated for 20 minutes, resulting in the development of a blue color. The color development is stopped with the addition of stop solution and the color is changed to yellow and measured spectrophotometrically at 450 nm. The concentration of TSH is directly proportional to the color intensity of the test sample.

REAGENTS AND MATERIALS PROVIDED:

1. Antibody-coated microtiter wells
2. Reference standard, Ready to use (0, 1, 2.5, 5, 10, 25 ng/mL), 0.6mL/vial
3. Enzyme Conjugate Reagent, 12 mL
4. TMB color reagent (ready to use) , 12 mL
5. 20X Wash buffer, 20 mL
6. Stop solution (2N HCl), 6mL
7. Instructions

MATERIALS REQUIRED, BUT NOT PROVIDED

1. Precision pipettes: 50uL, 100uL, 200uL, and 1.0mL
2. Disposable pipette tips
3. Vortex mixer or equivalent
4. Absorbent paper or paper towel
5. Graph paper
6. Microtiter plate reader

SPECIMEN COLLECTION AND PREPARATION

Serum should be prepared from a whole blood specimen obtained by acceptable medical techniques. This kit is for use with serum/plasma samples only.

STORAGE OF TEST KIT AND INSTRUMENTATION

Note of Caution: Immediately after receiving the kit all standards, if not used, should be kept at -20°C. Unopened test kits should be stored at 2-8°C. The microtiter plate should always be kept in a sealed bag with desiccants to minimize exposure to damp air at room temperature. Opened test kits will remain stable until the expiration date shown, provided it is stored as prescribed above. Do not leave any reagents at room temperature more than 3 hours.

A microtiter plate reader with a bandwidth of 10nm or less, with a bandwidth of 10nm or less and an optical density range of 0-3 OD or greater at a 450nm wavelength is acceptable for use in absorbency measurement.

REAGENT PREPARATION

1. All reagents should be brought to room temperature (25-28°C) before use.
2. The standards should be kept frozen at -20°C if not used immediately.
3. Dilute wash buffer, desired amount with distilled water (1 part with 19 parts). The buffer is stable for 1-3 months, if stored at 4-8°C.

ASSAY PROCEDURE

One must follow accurately these steps to ensure correct results. Use clean pipettes and sterile, disposable tips:

1. Secure the desired number of coated wells in the holder.
2. Dispense 100 µl of standards, specimens, and controls into appropriate wells.
3. Dispense 100 µl of Enzyme Conjugate Reagent into each well.
4. Thoroughly mix for 30 seconds. It is very important to have complete mixing at this step.
5. Incubate at 37°C for 3 hours in a sealed container or use zip-lock bag (provided).
6. Remove the incubation mixture by decanting the plate contents into a waste container.
7. Rinse and decant the microtiter wells five (5) times with wash buffer.
8. Strike the wells sharply onto absorbent paper or paper towels to remove all residual water droplets.
9. Dispense 100 µl of TMB solution into each well. Gently mix for 10 seconds.
10. Incubate at room temperature for 20 minutes.
11. Stop reaction by adding 50 µl (one drop) of stop solution, 2N HCl to each well.
12. Gently mix for 30 seconds. It is important to observe a color change from blue to yellow.
13. Read optical density at 450 nm with a microtiter well reader.

Important note: The wash step is critical. Insufficient washing will result in poor precision and falsely elevated absorbance readings

CALCULATION OF RESULTS

Calculate the mean absorbency value (A₄₅₀) for each set of reference standards, specimens, controls and test samples. Construct a standard curve by plotting the mean absorbency obtained from each reference standard against its concentration in ng/ml on graph paper, with absorbency values on the vertical or Y axis, and concentration on the horizontal or X axis. Use the mean absorbency values for each specimen to determine the corresponding concentration of TSH in ng/ml from the standard curve.

EXPECTED VALUES AND SENSITIVITY

The minimal detectable concentration of Rodent TSH by this assay is estimated to be 0.2 ng/ml. and the normal and experimental values should be established in your own laboratory. Each lab must follow good lab practice and maintain proper documentation.

REFERENCES

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Rodent TSH ELISA Test Kit**R & D use only**35325 Fircrest Street, Newark, CA 94560-1003 * Phone (800) 745-0843 * (510) 745-0844 * Fax (510) 745-0977

Quality Control Data:

It is highly recommended that each laboratory must establish their own internal controls and normal reference values for desired age, sex and physiological parameters.

A typical standard curve (illustration only) for Rodent TSH is given below:

Standard ng/mL	OD at 450nm
0	0.09
1	0.19
2.5	0.54
5	0.99
10	1.83
25	2.67

ELISA Performance Characters

Precision: Inter and Intra assay variation (CV) were determined from three different pooled serum samples in three different experiments.

Inter-assay variation	Set1: CV= 4.5% (N=10)	Set2: CV= 5.4 % (N=10)	Set3: CV= 4.2 % (N=10)
Intra-assay variation	Set1: CV= 4.8% (N=10)	Set2: CV= 5.8 % (N=10)	Set3: CV= 5.4 % (N=10)

Sensitivity: The lowest level detectable in this assay is 0.2ng/mL of serum or plasma

Specificity: The Rodent TSH ELISA system utilizes monoclonal antibody and high affinity polyclonal antibody to rodent TSH. The cross reactivity to other pituitary gonadotropins (Rodent LH, FSH) is not detectable under the conditions of the assay system.

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