Neospora caninum is a protozoon that was originally described as a parasite in dogs, in which it causes myositis and encephalitis. Bovine neosporosis is now recognised as a major cause of spontaneous abortion in cattle. It is highly suspected on 20% of the farms with repeated abortions and a cow that is seropositive for Neospora caninum has a threefold greater risk of aborting than a cow that is Neospora-negative. Neospora caninum is responsible for 21% of spontaneous abortions occurring in an individual animal. This percentage rises to 33% for the herd as a whole. Vertical transmission is the rule (at least 80% of the calves born to seropositive cows are infected). Serotesting before the calf’s first colostrum intake will reveal prenatal infection.

Use of the kit
The kit is designed to follow serological status of sera or milks

Reliable Results
The use of monoclonal antibody as conjugate ensures excellent specificity and very reliable results. The use of monoclonal antibodies to purify SRS2 from Neospora caninum protein on the plate also makes it possible to obtain an excellent specificity

Ease-of-Use
Minimal hands-on-time
Room temperature incubation
Results available in 140 minutes for single or batch testing

EIA Procedure

1- Microplate coated with monoclonal antibody and Neospora caninum purified SRS2 protein.

2- Add samples and positive control. Incubate 1 hour at 21°C+/-3°C. Wash

3- Add conjugate. Incubate 1 hour at 21°C+/-3°C. Wash

4- Add chromogen (TMB). Wait 10 minutes. Add stop solution. Read at 450 nm
The performance of the BIO K 192 kit (immunocapture test) was compared with BIO K 218 ELISA kit (blocking test) test on 270 blood serum samples. The results of these comparisons are shown in Graph 1. 1 corresponds to the value obtained with the kit’s positive reference serum (S/P).

Graph 1

Concordance between the two tests: Kappa = 0.83
The concordance between the two tests is considered excellent. Landis et Koch, The measurement of observer agreement for categorical data Biometrics 1977, 33, 159-74
269 serum and 269 milk samples taken from the same animals were tested using the BIO K 192 kit. These samples came from twenty-seven Belgian farms. Their optical density readings were divided by the optimal density reading for the kit’s reference serum (S/P). Frequency histograms were then plotted for the blood sera (Graph 2) and milk samples (Graph 3).
The frequency histograms generated for the 269 milk and 269 blood serum samples reveal that the thresholds must be set preferentially at 20% of the kit’s reference serum’s signal. Graph 4 shows the correlation between the serological findings yielded by the blood sera and those yielded by the milk samples.

Graph 4

Concordance between the two tests: Kappa = 0.85
The concordance between the two tests is considered excellent.
Landis et Koch, The measurement of observer agreement for categorical data
Biometrics 1977, 33, 159-74
Composition of the kit

BIO-X *NEOSPORA CANINUM* ELISA KIT : BIO K 192

<table>
<thead>
<tr>
<th></th>
<th>BIO K 192/2</th>
<th>BIO K 192/5</th>
</tr>
</thead>
<tbody>
<tr>
<td>Microplates</td>
<td>2 (96 tests)</td>
<td>5 (240 tests)</td>
</tr>
<tr>
<td>Washing solution</td>
<td>1 X 100 ml (20 X)</td>
<td>1 X 250 ml (20 X)</td>
</tr>
<tr>
<td>Dilution buffer</td>
<td>1 X 50 ml (5 X)</td>
<td>1 X 100 ml (5 X)</td>
</tr>
<tr>
<td>Conjugate</td>
<td>1 X 0.5 ml (50 X)</td>
<td>1 X 1.4 ml (50 X)</td>
</tr>
<tr>
<td>Positive serum</td>
<td>1 X 0.5 ml (1 X)</td>
<td>1 X 0.5 ml (1 X)</td>
</tr>
<tr>
<td>Single component TMB</td>
<td>1 X 25 ml (1 X)</td>
<td>1 X 55 ml (1 X)</td>
</tr>
<tr>
<td>Stopping solution</td>
<td>1 X 15 ml (1 X)</td>
<td>1 X 30 ml (1 X)</td>
</tr>
</tbody>
</table>

One year stability between +2°C and +8°C

Bibliography

NEOSPORA CANINUM ELISA KIT

ELISA kit for serodiagnosis of Neospora caninum
Indirect test for blood sera and milk
Diagnostic test for cattle

I - INTRODUCTION

Neospora caninum is a protozoon that was originally described as a parasite in dogs, in which it causes myositis and encephalitis. Bovine neosporosis is now recognised as a major cause of spontaneous abortion in cattle. It is highly suspected on 20% of the farms with repeated abortions and a cow that is seropositive for Neospora caninum has a threefold greater risk of aborting than a cow that is Neospora-negative. Neospora is responsible for 21% of spontaneous abortions occurring in an individual animal. This percentage rises to 33% for the herd as a whole. Vertical transmission is the rule (at least 80% of the calves born to seropositive cows are infected). Serotesting before the calf’s first colostrum intake will reveal prenatal infection.

II – PRINCIPLE OF THE TEST

The test uses 96-well microtitration plates sensitised by a purified Neospora caninum protein. The plate's odd columns (1, 3, 5, 7, 9 and 11) contain the purified protein, whereas the even columns (2, 4, 6, 8, 10 and 12) contain a control antigen. We thus have a genuine negative control. Using such a control reduces the number of false positives considerably.

The test blood sera or milks are diluted in the buffer for dilution. The plate is incubated and washed, then the conjugate, a peroxidase-labelled anti-bovine IgG1 monoclonal antibody, is added to the wells. The plate is then incubated a second time at 21°C +/- 3°C washed again and the chromogen tetramethylbenzidine (TMB) is added. This chromogen has the advantages of being more sensitive than the other peroxidase chromogens and not being carcinogenic. If specific anti-Neospora caninum immunoglobulins are present in the test sera or milks the conjugate remains bound to the microwell that contains the protozoon and the enzyme catalyses the transformation of the colorless chromogen into a pigmented compound. The intensity of the resulting blue colour is proportionate to the titre of specific antibody in the sample. The signal read off the negative control microwell is subtracted from that of the positive microwell sensitised by the protozoon protein.

III - COMPOSITION OF THE KIT

- **Microplates**: 96-well microtitration plates (6 strips of 16 wells). The odd columns (1, 3, 5, 7, 9 and 11) are sensitised by purified protein from Neospora caninum and the even columns (2, 4, 6, 8, 10 and 12) by the control antigen.
- **Washing solution**: One bottle of 20x concentrated washing solution. The solution crystallises spontaneously when cold. If only part of the solution is to be used, bring the bottle to 21°C +/- 3°C until all crystals have disappeared. Mix the solution well and remove the necessary volume. Dilute the buffer 1:20 with distilled or demineralised water.
**IV - ADDITIONAL MATERIALS AND EQUIPMENT REQUIRED**

Distilled water, graduated cylinders, beakers, plastic tubes, tube rack, dispenser tips, reagent reservoir for multichannel pipettes, lid, adhesive for microplates, graduated automatic (mono- and multichannel) pipettes, microplate reader, and microplate washer and shaker (optional)

**V - PRECAUTIONS FOR USE**

- This test may be used for “in vitro” diagnosis only. It is strictly for veterinary use.
- The reagents must be kept between +2°C and +8°C. The reagents cannot be guaranteed if the shelf-life dates have expired or if they have not been kept under the conditions described in this insert.
- The concentrated wash solution and dilution buffer may be stored at room temperature. Once diluted, these solutions remain stable for six weeks if kept between +2°C and +8°C.
- Unused strips must be stored immediately in the aluminium envelope, taking care to keep the desiccant dry and the envelope’s seal airtight. If these precautions are taken, the strips’ activity can be conserved up to the kit’s shelf-life date.
- Do not use reagents from other kits.
- The quality of the water used to prepare the various solutions is of the utmost importance. Do not use water that may contain oxidants (e.g., sodium hypochlorite) or heavy metal salts, as these substances can react with the chromogen.
- Discard all solutions contaminated with bacteria or fungi.
- The stop solution contains 1 M phosphoric acid. Handle it carefully.
- All materials and disposable equipment that come in contact with the samples must be considered potentially infectious and be disposed of in compliance with the legislation in force in the country.
- To guarantee the reliability of the results, one must follow the protocol to the letter. Special care must be taken in observing the incubation times and temperatures, as well as measuring the volumes and dilutions accurately.

**VI – PROCEDURE**

1. Bring all components to 21°C +/- 3°C before use. Remove the microplate from its wrapper.
2. For blood serum samples, place 1 ml dilution buffer, prepared as instructed in the « Composition of the Kit » section, in 5- or 10-ml hemolysis tubes. Add 10 µl of the serum samples to each of these tubes and shake briefly on a mechanical agitator.
3. Prepare milks in the following way: centrifuge for 20 minutes at 4000 g. Through the superior layer of the cream, using a glass Pasteur pipette, collect the intermediate liquid, making sure you don’t touch the...
underlying cellular pellet. Dilute skimmed milks to ¼ in the dilution solution (1 part of milk + 3 parts of dilution solution).

4- Dilute the positive serum of the kit to 1/100.
5- Distribute 100 µl of the samples per well, respecting the following pattern: positive serum: wells A1 and A2, sample 1: wells B1 and B2, sample 2: wells C1 and C2 etc… Incubate the plate at 21°C +/- 3°C for one hour. Cover the plate with a lid.
6- Rinse the plate with the washing solution, prepared as instructed in the "Composition of the Kit" section, as follows: empty the microplate of its contents by flipping it over sharply above a sink. Tap the microplate upside down against a piece of clean absorbent paper to remove all the liquid. Fill the used wells with the washing solution using a squeeze bottle or by plunging the plate in a vessel of the right dimensions, then empty the wells once more by turning the plate over above a sink. Repeat the entire operation two more times, taking care to avoid the formation of bubbles in the microwells. After the plate has been washed three times go on to the next step. An automatic plate washer may also be used, but in this case particular care must be taken to avoid any contact between the needles and the bottom of the wells to prevent any damage of the reagent layer.

7- Dilute the conjugate 1:50 in the dilution buffer (for example, for one plate dilute 250 µl of the conjugate stock solution in 12,250 ml of diluent). Add 100 µl of the dilute conjugate solution to each well. Incubate for 1 hour at 21°C +/- 3°C. Cover the plate with a lid.
8- Wash the plate as described in step 6 above.
9- Add 100 µl of the chromogen solution to each well on the plate. The chromogen solution must be absolutely colourless when it is pipetted into the wells. If a blue colour is visible, this means that the solution in the pipette has been contaminated.
10- Incubate for 10 minutes at 21°C +/- 3°C protected from the light and uncovered. This time is given as a guideline only, for in some circumstances it may be useful to lengthen or shorten the incubation time.
11- Add 50 µl of stop solution per microwell.
12- Read the optical densities in the microwells using a plate reader and a 450 nm filter. Results must be read fairly soon after the stopping solution has been added since the chromogen may crystallize in wells with strong signals and thereby distort the data.

VII – INTERPRETING THE RESULTS

Subtract from each value recorded for the odd columns the signal of the corresponding negative control well and write down the result. In performing this calculation, allow for any negative values that may exist. Carry out the same operations for the column corresponding to the positive control.

The test can be validated only if the positive serum yields a difference in optical density at 10 minutes that is greater than 0,800

Divide the signal read for each sample well by the corresponding positive control serum signal and multiply this result by 100 to express it as a percentage.

\[
\text{Val} = \frac{\Delta \text{OD Sample}}{\Delta \text{OD positive}} \times 100
\]

Using the following table, determine each serum or milk status (Positive, doubtful or negative).

<table>
<thead>
<tr>
<th>Val</th>
<th>0</th>
<th>+/-</th>
<th>+</th>
</tr>
</thead>
<tbody>
<tr>
<td>Val</td>
<td>&lt;</td>
<td>10 %</td>
<td>&lt; Val</td>
</tr>
</tbody>
</table>

VIII – ORDERING INFORMATION

BIO-X Neospora caninum ELISA KIT:

| 2x48 tests | BIO K 192/2 |
| 5x48 tests | BIO K 192/5 |