

**FOR *in vitro* USE ONLY**

**Reconstituted *in vitro* translation kit**

**PURESYSTEM<sup>TM</sup> S-S**

## **Instruction Manual**

<b>96</b>	<b>PURE3048S</b>
<b>standard</b>	<b>PURE3030S</b>
<b>mini</b>	<b>PURE3004S</b>

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**IMPORTANT NOTICE to Purchaser**

- This kit cannot be used in human subjects, in clinical trials or for diagnostic purposes involving human subjects.
- Contents of this kit are subject to change without notice.

**Safety Instructions**

Please note that Sol. A and Sol. B contains <1% DTT (Dithiothreitol). No other hazardous substances are included in reportable quantities. The synthesized protein may have strong toxicity or show pathogenic properties. To insure your safety, please take all the necessary precautions such as using the safety cabinet or protecting your eyes and skin by use of eyewear and gloves at all times. If your skin comes in direct contact with any of the reagents, or if you have swallowed any reagents by mistake, please obtain appropriate medical assistance. Please also insure to inactivate all unused reagents by autoclave or dry-heat sterilization before disposal. If your institution or country holds its own laws or regulations concerning experimental procedures, material transfer, export and disposal of the materials, please follow the instructions as stated.

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## 1. Quick Protocol for Protein Synthesis Reaction

1. Thaw Sol. A and Sol. B on ice.
2. Assemble in new reaction tube(s) as follows;

	96 / mini	standard
<input type="checkbox"/> Nuclease-free water	μl	μl
<input type="checkbox"/> Sol. A	25 μl	500 μl
<input type="checkbox"/> Sol. B	10 μl	200 μl
<input type="checkbox"/> Template DNA	μl* <sup>1</sup>	μl* <sup>2</sup>
Total	50 μl	1000 μl

\*1; 1 pmol PCR product or 0.5 μg plasmid DNA

\*2; 20 pmol PCR product or 10 μg plasmid DNA

3. Mix gently and centrifuge briefly.
4. Incubate the reaction(s) at 37°C for 1 hour.
5. Stop the reaction(s) by placing the tube(s) on ice.
6. Analyze the synthesized protein(s).

## 2. Kit Components

Please check that your kit contains the reagents listed below.

### 2-1. PURESYSYSTEM™ S-S 96 (Prod.#PURE3048S)

Each kit contains sufficient reagents for 96 x 50 µl reactions.

<b>Label (Tube Color)</b>	<b>Amount</b>
Sol. A (Green)	96 x 25 µl
Sol. B (Orange)	96 x 10 µl
Univ. Primer (Clear)	2 x 500 µl

### 2-2. PURESYSYSTEM™ S-S standard (Prod.#PURE3030S)

Each kit contains sufficient reagents for 3 x 1000 µl reactions.

<b>Label (Tube Color)</b>	<b>Amount</b>
Sol. A (Green)	3 x 500 µl
Sol. B (Orange)	3 x 200 µl
Univ. Primer (Clear)	1 x 80 µl

### 2-3. PURESYSYSTEM™ S-S mini (Prod.#PURE3004S)

Each kit contains sufficient reagents for 8 x 50 µl reactions.

<b>Label (Tube Color)</b>	<b>Amount</b>
Sol. A (Green)	8 x 25 µl
Sol. B (Orange)	8 x 10 µl
Univ. Primer (Clear)	1 x 80 µl

- Sol. A and Sol. B must be thawed just before use on ice and avoided multiple freeze-thaw cycles.
- Univ. Primer is a forward primer for second-step PCR and its concentration is 2 µM.
- Template DNA of alkaline phosphatase (AP) is available for synthesis of control protein containing disulfide bonds. The option kit (AP template & substrate, Prod.#PUREAP01) includes template DNA of AP and substrate for the enzyme. We strongly recommend you to synthesize both protein of interest and AP together in order to ascertain the experimental procedures.

## 3. Storage Conditions

Store all the components in the kit at -80°C. Avoid multiple freeze-thaw cycles.

## 4. Description

### 4-1. Background of PURESYSTEM Technology

PURESYSTEM is a novel reconstituted *in vitro* translation system which consists of about 30 purified enzymes necessary for transcription, translation and energy recycling.\* In PURESYSTEM, all the factors for transcription and translation are tagged with a hexahistidine; 3 initiation factors (IF1, IF2, IF3), 3 elongation factors (EF-G, EF-Tu, EF-Ts), 3 release factors (RF1, RF2, RF3), ribosome recycling factor, 20 aminoacyl-tRNA synthetases, methionyl-tRNA formyltransferase and T7 RNA polymerase. The reagents also contains *E. coli* 70S ribosome, amino acids, NTPs, *E. coli* tRNA and energy recycling system, so the target protein can be synthesized just by addition of the template DNA to the reaction.

\* Shimizu Y., *et al.*, (2001) *Nature Biotechnology*, vol.19, p.751-755.

### 4-2. PURESYSTEM S-S Kit

PURESYSTEM S-S is the world's first *in vitro* translation system which allows the synthesized protein to form disulfide bond(s).

#### Disulfide Bond Formation

PURESYSTEM S-S is especially designed to enable the formation of disulfide bond(s) of synthesized protein. This was previously believed to be impossible with conventional *in vitro* protein synthesis systems including *E. coli* S30 extract, wheat germ extract and rabbit reticulocyte lysate systems. The high purity and flexibility of PURESYSTEM enables the redox conditions of reaction mixture to be reproducible. PURESYSTEM S-S consists solely of defined components and does not contain any contaminants such as glutathione reductase and thioredoxin reductase which may cause redox reactions.

## **High Purity**

Because PURESYSTEM is a reconstituted system in which all the components are identified, the reagents in PURESYSTEM S-S are essentially free from unidentified components. Therefore, the synthesized product is hardly susceptible to unwanted degradation or modification.

## **4-3. Related Kits**

### **PURESYSTEM™ classic II**

PURESYSTEM classic II, the primary kit based on PURESYSTEM technology, enables synthesis reaction under reduced condition.

### **PURESYSTEM™ custom**

PURESYSTEM is a reconstituted system, so the reagent composition can be easily modified according to the purpose of experiment. This can be ordered as PURESYSTEM custom, which is made upon request.

### **PURESYSTEM™ Duo**

PURESYSTEM Duo is an introductory *in vitro* transcription/translation kit which contains both classic II (reduced condition) and S-S (oxidized condition) reagents.

## 5. Procedures

### 5-1. Preparation of Template DNA

#### 5-1-1. Considerations for Template DNA Construction

Both PCR product and plasmid DNA can be used as the template DNA for protein synthesis reaction in **PURESYSYSTEM**. The template DNA needs to contain the following;

- start codon (ATG)
- stop codon (TAG, TGA or TAA)
- T7 promoter sequence at upstream from the coding sequence
- Ribosomal binding site (RBS; SD sequence) at approximately 10 bp upstream from start codon
- Additional sequence of 6 bp or more at downstream from stop codon (for PCR product)
- Terminator sequence at downstream from stop codon (for plasmid DNA)

We recommend PCR product as the template DNA due to the following reasons;

- PCR product can be used without further purification.
- No need for time-consuming construction of plasmid DNA suitable for protein synthesis.
- Multiple templates can be prepared at a time.
- The coding sequence can be easily modified.  
eg) deletion, point mutation, division into several domains and addition of tag sequence.

#### 5-1-2. Generation of the Template DNA by 2-step PCR

The template DNA can be generated by 2-step PCR (Figure 1). In the first (1<sup>st</sup>) PCR, the adapter sequence is added at 5'-terminus of the gene of your interest, and all the regulatory sequences are added in the second (2<sup>nd</sup>) PCR. It is possible to amplify the template DNA by 1-step PCR using forward primer containing 5'-terminal region of the coding sequence and all regulatory sequences.

1. Design the forward and reverse primers as shown in Table 1.

2. Perform 1<sup>st</sup> PCR from DNA containing the gene of interest using these primers and high-fidelity polymerase.
3. Analyze the 1<sup>st</sup> PCR product by agarose gel electrophoresis. When detected as a single band, the 1<sup>st</sup> PCR reaction mix can be used in the second step without further purification. If any additional bands are detected, please optimize the PCR conditions or excise the band of the product from agarose gel.
4. Perform 2<sup>nd</sup> PCR from the 1<sup>st</sup> PCR product using universal primer included in the kit and reverse primer used at the 1<sup>st</sup> PCR.
5. Analyze the 2<sup>nd</sup> PCR product by agarose gel electrophoresis. The 2<sup>nd</sup> PCR product will be approximately 70 bp longer than the 1<sup>st</sup> PCR product, and this can be used for protein synthesis reaction without further purification.

**Table 1. Primer List**

primer	sequence
Forward primer	5'-AAGGAGATATACCA-ATG-N <sub>14-20</sub> -3'
Reverse primer	5'-TATTCA-TTA-N <sub>14-20</sub> -3'
Universal primer (Univ. Primer)	5'-GAAATTAATACGACTCACTATAGGGAGACCACAACGGTTTCC CTCTAGAAATAATTTTGTTTAACTTTAAGAAGGAGATATACCA-3'

### 5-1-3. Considerations for plasmid DNA

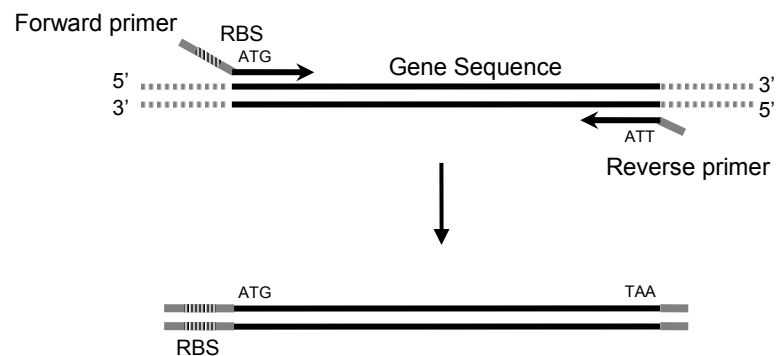
Plasmid DNA can also be used as the template DNA for **PURESYSYSTEM**. The plasmid DNA will need to fulfill the following elements and structural features;

- Any commercially available expression vectors can be used which contains T7 promoter sequence, RBS (approximately 10 bp upstream from the initiation codon) and terminator sequence.
- Some commercially available plasmid purification kits are not suitable for preparation of template DNA for **PURESYSYSTEM**. Please contact technical service for details.
- The plasmid DNA needs to be dissolved in RNase-free water or buffer without EDTA, in order to avoid inhibition of protein synthesis reaction.
- RNA needs to be removed. When RNase is used for the removal of RNA, complete inactivation of RNase is necessary by Phenol treatment or by similar

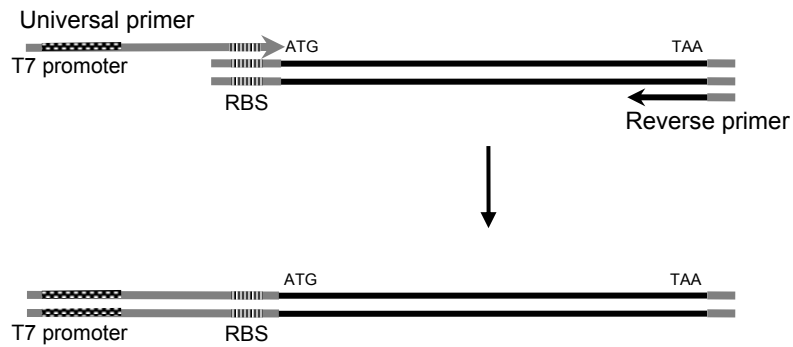
inactivation treatments.

- Quick purification method described in this manual cannot be used when the target gene is introduced to expression vector with a histidine-tag sequence.

### 1<sup>st</sup> Step PCR



### 2<sup>nd</sup> Step PCR



**Figure 1. Generation of template DNA by 2-step PCR**

## 5-2. Protein Synthesis Reaction

### 5-2-1. Precautions concerning reagents and experimental procedures

If you are unfamiliar with an *in vitro* translation, we strongly recommend you to check the experimental procedures by using negative control (i.e., add water instead of template DNA) and positive control (alkaline phosphatase), which is available as an option kit (see page 4 and 19 for ordering information). In order to avoid nuclease contamination, please use Nuclease-free tubes and tips, and wear gloves and mask.

### 5-2-2. Procedures

1. Thaw the necessary number of Sol. A and Sol. B on ice.

Please avoid multiple freeze-thaw cycles, because some enzymes in the reagents may be inactivated.

2. Assemble in new reaction tube(s) as follows;

	mini / 96	standard
<input type="checkbox"/> Nuclease-free water	μl	μl
<input type="checkbox"/> Sol. A	25 μl	500 μl
<input type="checkbox"/> Sol. B	10 μl	200 μl
<input type="checkbox"/> Template DNA	μl* <sup>1</sup>	μl* <sup>2</sup>
Total	50 μl	1000 μl

\*1; 1 pmol PCR product or 0.5 μg plasmid DNA

\*2; 20 pmol PCR product or 10 μg plasmid DNA

When labeled amino acid is incorporated into synthesized protein, add the labeled amino acid just before protein synthesis. For example, add 1 μl of 0.4 MBq/μl [<sup>35</sup>S]methionine before addition of template DNA.

3. Mix gently and centrifuge briefly.
4. Incubate the reaction(s) at 37°C for 1 hour.
5. Stop the reaction by placing the tube(s) on ice.

The reaction mixture is now ready to be analyzed. (e.g., purification, electrophoresis, enzyme assay, binding assay).

Some proteins may require couple of hours for correct folding to take place after synthesis reaction. If correct folding is known to be crucial for your protein of interest, please place the tube in the incubator.

Refer to section 5-3 for Quick Purification.

If purification step is unnecessary, the reaction mixture may be directly used for your experiment.

### **5-3. Purification of Synthesized Protein - Quick Purification Protocol**

If your proteins are characterized and you are familiar with the purification methods for your proteins, please proceed with the methods of your choice.

If the purification methods of the synthesized protein are unknown and the molecular weight is less than 100 kDa, the protein may be purified by the quick purification method as follows.

#### **5-3-1. Additional Equipments necessary for Quick Purification**

Ni-NTA Agarose (QIAGEN)

Microcon YM-100 (Millipore)

Micro Bio-Spin Chromatography Column (BIO-RAD)

Vortex Mixer

#### **5-3-2. Procedure 1 (for 96 and mini)**

1. Add equal volume of H<sub>2</sub>O to the reaction.
2. Apply the diluted reaction mixture to Microcon YM-100.
3. Centrifuge at 1,500xg at 4°C for 30 minutes.
4. Transfer the permeate to a new tube (preferably 2 ml round-bottom tube).
5. Add 0.1 vol. of Ni-NTA Agarose.
6. Vortex for 1 hour at 4°C vigorously.
7. Apply the mixture to the Micro Bio-Spin Chromatography Column.
8. Centrifuge at 1,500xg at 4°C for 1 minute.
9. Recover the permeate.

#### **5-3-3. Procedure 2 (for 96 and mini)**

This procedure is recommended when the synthesized protein could be purified according to the Procedure 1.

1. Add equal volume of H<sub>2</sub>O to the reaction.
2. Transfer the diluted reaction mixture to a new tube (preferably 2 ml round-bottom tube).
3. Add 0.1 vol. of Ni-NTA Agarose.

4. Vortex for 1 hour at 4°C vigorously.
5. Apply the mixture to Microcon YM-100.
6. Centrifuge at 1,500xg at 4°C for 30 minutes.
7. Recover the permeate fraction.

#### **5-3-4. Procedure 3 (for standard)**

1. Divide the reaction mixture into 200 µl-aliquot and apply each aliquot to Microcon YM-100.
2. Centrifuge at 1,500xg at 4°C for 30 minutes.
3. Pool the permeate in a new 2 ml round-bottom tube.
4. Add 0.1 vol. of Ni-NTA Agarose.
5. Vortex for 1 hour at 4°C vigorously.
6. Apply the mixture to the Micro Bio-Spin Chromatography Column.
7. Centrifuge at 1,500xg at 4°C for 3 minutes.
8. Recover the permeate.

#### **5-4. Analysis of Synthesized protein**

Synthesized proteins can be detected and analyzed by methods such as SDS-PAGE followed by gel staining or western blotting and enzyme activity assay. In this section, a general method for SDS-PAGE and gel staining is shown.

If protein synthesis reaction is carried out in the presence of radiolabeled amino acids, the product can be analyzed by fluorography.

1. Mix 1-10  $\mu$ l of samples with SDS-PAGE sample buffer.
2. Load the samples to SDS-PAGE gel and carry out electrophoresis.
3. Stain the gel by Coomassie Brilliant Blue, Silver, or Fluorescent dye such as SyproOrange.

The yield of the target product may be dependent on each protein. If the yield is low, please concentrate the samples by methods such as trichloroacetic acid and acetone. Depending on the staining method or kit, tRNA band may be detected around 20-30 kDa. The background can be reduced by treatment with RNase after synthesis reaction. For example, add 3.75  $\mu$ g of RNaseA in 50  $\mu$ l of reaction and incubate at 37°C for 15 min.

## 6. Synthesis and Enzyme Assay for AP

Template DNA of alkaline phosphatase (AP) is available for synthesis of control protein containing disulfide bonds. The option kit (AP template & substrate, Prod.#PUREAP01) includes template DNA of AP and substrate for the enzyme. We strongly recommend you to synthesize both protein of interest and AP together in order to ascertain the experimental procedures.

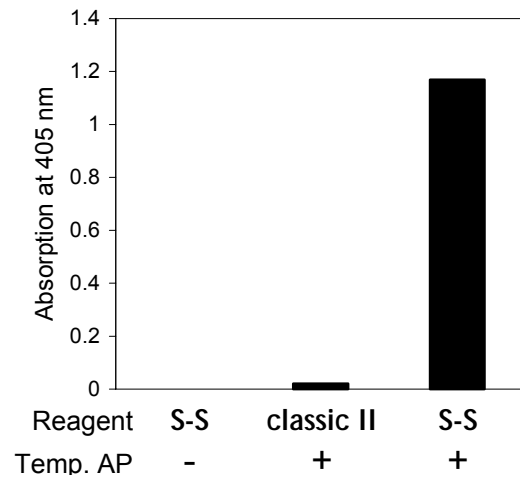
1. Thaw Sol. A, Sol. B and Temp. AP on ice.
2. Assemble in a new reaction tube as follows;

<input type="checkbox"/> Nuclease-free water	12.5 $\mu$ l	
<input type="checkbox"/> Sol. A	25 $\mu$ l	
<input type="checkbox"/> Sol. B	10 $\mu$ l	
<input type="checkbox"/> Temp. AP	2.5 $\mu$ l	(0.5 $\mu$ g)
<hr/>		
Total	50 $\mu$ l	

3. Mix gently and centrifuge briefly.
4. Incubate the reaction at 37°C for 1 hour.
5. Stop the reaction by placing the tube on ice.
6. Mix Sub. AP and Buf. AP to dissolve the substrate.
7. Assemble in 96 well plate as follows;

<input type="checkbox"/> Reaction mixture	5 $\mu$ l
<input type="checkbox"/> Substrate solution	95 $\mu$ l

8. Incubate at 37°C for 30 minutes.
9. Read absorbance at 405 nm by spectrophotometer.



**Figure 2 Enzyme activity of synthesized AP**

## 7. Troubleshooting

### 7-1. Preparation of Template DNA

#### No product at the 1<sup>st</sup> step PCR

Primer is not optimal.

→ Redesign the primer.

Condition is not optimal.

→ Check reaction conditions such as annealing temperature, extension time and number of cycles.

→ Perform the PCR reaction in the presence of additives such as DMSO.

#### No product at the second-step PCR

Primer is not optimal.

→ Redesign the primer.

→ Prepare the template DNA by 1 step PCR using the long primer as below.

5'-GAAATTAATACGACTCACTATAGGGAGACCACAACGGTTTCC  
CTCTAGAAATAATTTTGT'TTAAC'TTTAAGAAGGAGATATACCA-ATG-N<sub>14-20</sub>-3'

Condition is not optimal.

→ Check reaction conditions such as annealing temperature, extension time and number of cycles.

→ Perform the PCR reaction in the presence of additives such as DMSO.

### 7-2. Protein Synthesis by PURESYSTEM

#### No control is synthesized.

Some components in the kit are inactivated.

→ Store the reagents in the kit at -80 °C and avoid multiple freeze-thaw cycles.

The reaction is contaminated with nuclease.

→ Wear gloves and mask and use nuclease-free tubes and tips.

#### Control is synthesized, but the target is not.

DNA sequence of the template DNA is incorrect.

→ Check the sequence of the template DNA.

Purity and/or concentration of the template DNA is not optimal.

→ Check purity and concentration of the template DNA.

The reaction is contaminated with nuclease.

→ Wear gloves and mask and use nuclease-free tubes and tips.

Transcript may have formed a rigid secondary structure.

Introduce mutations in the sequence to avoid secondary structure formation.

Insert a tag coding sequence at the 5' terminus of the target sequence.

Products (transcripts and/or polypeptides) inhibit protein synthesis reaction.

Synthesize control protein in the same reaction mixture and check the activity of AP.

The condition for protein synthesis is not optimum for some sort of proteins.

The optimum condition such as redox condition, concentration of chaperones, and reaction temperature may vary according to the each protein.

**The target protein is synthesized, but the yield or activity is very low.**

Purity and/or concentration of the template DNA is not optimal.

Check purity and concentration of the template DNA.

The reaction is contaminated with nuclease.

Wear gloves and mask and use nuclease-free tubes and tips.

Transcript may form a rigid secondary structure.

Introduce mutations into the sequence to avoid secondary structure formation.

The condition for protein synthesis is not optimum for some sort of proteins

The optimum condition such as redox condition, concentration of chaperones, and reaction temperature may vary according to each protein.

**The target protein is synthesized, but it is insoluble.**

Products has formed aggregates.

Protein solubility is improved when protein synthesis reaction is performed at temperatures lower than 37°C.

The condition for protein synthesis is not optimum for some sort of proteins

The optimum condition such as redox condition, concentration of chaperones, and reaction temperature may vary according to each proteins.

## 8. Technical Service

For information about PURESYSTEM technology and kits (including custom kit)

**info@biocomber.co.jp**

For our latest information

**www.biocomber.co.jp**

## 9. Related Products

<b>Product</b>	<b>Pack size</b>	<b>Prod. No.</b>
<b>PURESYSTEM classicII 96</b>	96 x 50 µl reactions	PURE2048C
<b>PURESYSTEM classicII standard</b>	3 x 1 ml reactions	PURE2030C
<b>PURESYSTEM classicII mini</b>	8 x 50 µl reactions	PURE2004C
<b>PURESYSTEM CHAPERONE DnaK set</b>	40µl DnaK, 20µl DnaJ, 20µl GrpE	PCDS01
<b>PURESYSTEM CHAPERONE GroE set</b>	20µl GroEL, 20µl GroES	PCGS01
<b>PURESYSTEM Duo</b>	<b>classic II</b> (4 x 50 µl reactions) <b>S-S</b> (4 x 50 µl reactions)	PURE4004D
<b>AP template &amp; substrate</b>	3 µg template DNA 1000 µl substrate solution	PUREAP01
<b>Universal Primer</b>	2 µM x 500 µl	PURER001

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